Stevia (*Stevia rebaudiana* B.) as a medicinal plant and its antioxidant and lipid-modulating effects – with focus on ruminant production. A review

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Abstract

Penchev, P., Enchev, S., Nenova, R., Ilieva, Y., Ivanova, D. & Schreiner, M. (2025). Stevia (*Stevia rebaudiana* B.) as a medicinal plant and its antioxidant and lipid-modulating effects – with focus on ruminant production. A review. *Bulg. J. Agric. Sci.*, *31*(3), 595–604

A comprehensive literature search was conducted, and the expertise and achievements with stevia and some other medicinal plants of our plant and animal breedeing teams were taken in consideration. The objective to summarize the background of the medicinal applications of stevia was generally fuilfilled, the found available sources treating the effects on ruminant products is limited. Besides its direct benefits to consumers, indirect effect of its glycosides should be especially sought on production of ruminant origin, which is notorious for its saturated lipids. Except its immunomodulatory, anti-inflammatory, anti-oxidative and anti-tumor properties, this review implies stevia's capacity to modulate the anti-oxidant and nutritional qualities of foodstuffs. Not many proofs have been found for the influence of stevia on fatty acid composition of ruminant products, but its effects on rumen processes suggest application as a natural modulator of lipid quality via manipulation of rumen environment and functions. All this necessiates further in-depth research, but to date the available scientific evidence affords the presumption that it is worthwile to grow stevia for such purposes, justifying the peculiarities of its cultivation (vegetatively bred and maintained *in vitro*) and the efforts in the latitudes of Bulgaria, where it needs to be treated like an annual, not like a perennial plant as in its continent of origin.

Keywords: buffalo; medicinal plants; stevia, antioxidant; fatty acids

Introduction

The essential nutritional values of foodstuffs of ruminant origin are indisputable but still there is criticism against the mass consumption of milk and dairy products, because of the saturated nature of their lipids (Givens & Shingfield, 2006). In this context, according to FAO, saturated fatty acids should provide no more than 10% of total calories (FAO/ WHO, 2003).

It is noteworthy that milk contains highly desirable oleic acid and conjugated linoleic acid (CLA), with anti-cardio-

vascular, anti-carcinogenic, anti-diabetic and immunomodulating effects (Belury, 2002; Parodi, 2004). Mainly Δ 9-desaturase enzyme in the mammary gland (Bauman et al., 2006) and ruminal biohydrogenation (Shingfield et al., 2005; Kalač & Samková, 2010) are responsible for the expression of the effect of feeding on fatty acid profile of milk (Fernandes et al., 2007; Beyero et al., 2015) and meat (Daley et al., 2010; Nogoy et al., 2022).

Since 1970s, encapsulated/protected proteins and lipids have been used in cattle breeding achieving significant changes in the fatty acid profile in ruminant (Ashes et al., 1992; Scollan et al., 2003; Smith et al., 2010). Nevertheless, due to the hazards they cause to animals, plants and humans, the World Health Organization (WHO) encourages farmers to use medicinal plants and herbs to substitute or minimize their use, as a global trend that turns towards nature (Allam et al., 1999).

In the search for immunomodulatory agents to alleviate diseases, for many years the focus has been on finding medicinal plants that match the philosophy of 'Rasayana' and 'Ayurveda' and that possess such properties (Patwardhan et al., 1990; Diasio & LoBuglio, 1996).

In ruminants, herbal and medicinal plants have been also used to stimulate appetite and ruminal function resulting in health benefits for the animals (Cedillo et al., 2015; Salem et al., 2017). Moreover, phytogenic compounds have been used as alternatives of antibiotics, and to increase feed intake, blood chemistry and productivity (Matloup et al., 2017; Khattab et al., 2020).

In a study of Fahim et al. (2022) on buffalo cows, fennel was found to significantly increase the concentrations of trans-10, cis-12 C18:2, total CLA in milk without affecting other fatty acids. A dose of 75 g per capita per day of fennel or ginger improved nutrient digestibility and milk production of buffaloes.

Our experiment with a buffalo herd in Bulgaria supplemented a herbal preparation of the Piper genus (AyuFertin) showed a trend to affect the fatty acid profile of the milk of the treated buffaloes (Ilieva et al., 2022). The results of another our recent experiment with Curcumin C3 Complex on buffaloes (Penchev et al., 2022), showed increased concentration of the valuable vaccenic, rumenic acid and total trans-isomers in the raw milk after supplementation.-

Stevia rebaudiana, belonging to the sunflower family (Asteraceae), is a perennial herb native to the subtropical regions of South America that has been used for decades as a sweetener in many countries, predominantly in Latin America and East Asia (Lemus-Mondaca et al., 2012). The sweeteners produced from it have sensory and functional properties superior to the conventional sweeteners and have the perspective to prevail on the food market (Goyal et al., 2010). This is chiefly attributed to Stevioside (13-[(2-O-be-ta-D-glucopyranosyl-alpha-D-glucopy-ranosyl oxy] kaur-16-en-18-oic acid-beta-D-glucopyranosyl ester), which is one of the main glycosides in the green parts of Stevia rebaudiana, and which outstands with its safety, stability, low pH, high degree of sweetness, in the same time with low caloric value (Puri et al., 2011).

Also, the European Food Safety Authority (EFSA) approved the usage of steviol glycosides as a non-toxic alternative sweetener in food and beverages industry (Ilias et al.,

2021). Moreover, stevia was established to contain other components that have therapeutic properties including vitamins, minerals, essential amino acids, fatty acids, flavonoids, alkaloids, chlorophyll, xanthophyll, aromatic acids, chlorogenic acid, oligosaccharides, free sugars, and phytosterol (Muanda et al., 2011; Ahmad et al., 2020).

The first extraction of steviosides from stevia was in 1931, done by French chemists (Carakostas et al., 2008). In 1964 and 1968, stevia was commercially cultivated in Paraguay and Japan, respectively, and used extensively in commercial products. Japan legalized usage of stevia as a sweetener in the 1970s, banning and replacing all other chemical sweeteners. Since then, cultivation of the plant has expanded to other countries including China, Malaysia, Singapore, South Korea, Taiwan, Thailand, Paraguay, Brazil, USA, Canada and Europe (Midmore & Rank, 2002; Chatsudthipong & Muanprasatq, 2009).

In 1980, stevia was introduced in Bulgaria from Japan and USA and became a subject of research at the Agricultural Institute – Shumen. As a result of the scientific studies, stevia was introduced into culture *in vitro* and appropriate nutrient media were developed for its micropropagation and rooting (Slavova et al., 2003; Kikindonov, 2013; Uchkunov et al., 2015). At the Institute of Organic Chemistry Sofia was done the extraction and purification of the sweet substances from the dry leaves, and at the Institute of Nutrition at the Medical Academy was analyzed the sweet substances for toxicity and safety (Nikolova-Damyanova et al., 1994).

For more than 15 years, plants of several stevia genotypes were maintained under *in vitro* conditions. As a result of the selection, the first Bulgarian variety "Stella" was created and certified legally in 2016, now being bred vegetatively.

The soil and climate conditions in Bulgaria allow for the successful cultivation of *S. rebaudiana* B. Due to its high sensitivity to low temperatures, stevia is grown only as an annual plant (Kikindonov, 2013). At a later stage, in greenhouse conditions, it can be vegetatively bred – by means of green rhizomes obtained from planted for the purpose roots that had passed one growing season in field conditions, but it can also be developed from adapted regenerates from *in vitro* maintained branches, and under favorable conditions it is possible to obtain seeds from it (Mehmed et al., 2021).

Our studies have shown that the yield of glycosides, especially stevioside, strongly depends on the total biomass yield, which in turn depends on good agricultural practices for growing the plant (Kikindonov & Enchev, 2012; Mehmed & Enchev, 2020).

The stevioside and rebaudiside are the main steviol glycosides, which make up more than 95% of the total amount of sweet substances. Pure stevioside is about 200 times sweeter than sucrose, and rebaudiside A is about 250 times sweeter (Brandie, 2004). Although these substances are extremely sweet, they do not contain calories (Enchev et al., 2018; Tanova & Kaschieva, 2018) and are also resistant to high temperatures (up to 198°C) (Benford et al., 2006). They are non-fermentable and do not induce carbohydrate cravings, making them suitable for diabetics (Curi et al., 1986; Gregersen et al., 2004).

Objectives

With this review we aimed to summarize the background of the medicinal applications of *Stevia rebaudiana B*. and its derivates for oxidative stress and immune activity – in mammals as a whole, and in livestock in particular and their effects on the products thereof, with focus on foodstuffs of ruminant origin. In this context, the focus is on the water buffalo (*Bubalus bubalis*) and the produced milk.

Material and Methods

A profound comprehensive search of scientific literature was conducted in order to study the problem in focus. In the present review was used information from publications obtained on the Internet – exclusively Google Scholar but mostly Research Gate, PubMed, Scopus and Web of Science – as well as journal editions worldwide and also editions in Bulgaria. More specifically, the expertise and achievements with stevia of the team of plant breeders at Agricultural Institute – Shumen and also the research experience and experimental results with medicinal plants in ruminants of our team were taken in consideration, as well.

Results of Literature Review

In connection with the changes in the food industry and the modern trend for a healthy lifestyle, there is an increased scientific interest in natural immunomodulators and antioxidants in nature. Many countries are conducting their own research in order to discover natural, low-calorie sweeteners of plant origin that are not harmful to human body.

Functional benefits of stevia (in vitro investigations, laboratory animals and human)

The uses and applications of stevia are increasing both on national and international scale. It has been found to possess physiological activity that can be used in human healthcare. As a healthy replacement of sucrose, stevia has the ability to regulate blood pressure and blood sugar (Ferri et al., 2006; Gupta et al., 2016; Rotimi et al., 2018). In addition, it was revealed that stevia had the potential to positively affect blood sugar and insulin levels, urine sodium excretion, lipid profile and body weight (Savita et al., 2004).

Except for its exceptional sweetness, stevia is valuable because of its nutritional components, especially phytochemicals and polyphenols (Chu et al., 2000; Singh & Rao, 2005; Edeoga et al., 2005) contributing for its outstanding antioxidant properties (Chatsudthipong & Muanprasat, 2009; Ahmad et al., 2010).

According to the review by Thomas and Glade (Thomas & Glade, 2010) the antioxidant capacity of stevia is even greater than DL- α -tocopherol and green tea extract, due to the activity against free radicals and superoxides (Xi et al., 1998 a, b). It contains high concentrations of total polyphenols and flavonoids and prevents also lipid peroxidation and DNA strand excision (Ghanta et al., 2007). Another research also demonstrated the potential of *Stevia rebaudiana* in functional foods production or as microbial or therapeutic agent, finding a favorable correlation between phenolics and flavonoids on one hand and antioxidant capacity on the other (Lemus-Mondaca et al., 2018).

An experiment in human medicine showed that supplementation of stevia led to a decrease in blood glucose, alanine and aspartate transaminase and an increase in blood insulin level (Shivanna et al., 2013). Stevia also reduced malondialdehyde concentration in the liver and improved its antioxidant status through antioxidant enzymes. The authors result in optimized glucose tolerance and insulin sensitivity. The antioxidant capacity of stevia has found confirmation in other studies as well (Shukla et al., 2012; Bender et al., 2015).

The immune system support of stevia leaf extracts is expressed in significant antiviral and antimicrobial and antifungal activity towards a wide range of pathogenic bacteria, including enterohemorrhagic *Escherichia coli*, without affecting normal intestinal flora (Tomita et al., 1997; Ghosh et al., 2008). Furthermore, stevia inhibits the replication of rotavirus *in vitro* by blocking viral attachment to cells (Takahashi et al., 2001).

A very recent medicinal *in vitro* study reveals the ability of steviol glycosides (stevioside and rebaudioside A) to affect the expression of genes involved in glucose and lipid metabolism (Kurek et al., 2023).

A number of studies indicated the decreasing effect of stevia on triglyceride (Aghajanyan et al., 2017) and fatty acid synthesis due to reduced levels of acetyl-COA carboxylase mRNA (Park & Cha, 2010). It reduces LDL cholesterol, triglyceride, and increases HDL cholesterol (Sharma et al., 2009), and hence can be used as an instrument to alleviate cardiovascular diseases (Ilias et al., 2021).

Besides anti-oxidant activity, stevia glycosides have been

proven to posses wound healing, anti-viral, anti-inflammatory, anti-diabetic, and renal-protective properties, expressed in stimulation of the immune system functions like delayed type hypersensitivity response, macrophage phagocytosis, and B and T lymphocytes proliferation (Sehar et al., 2008; Cho et al., 2013). Isosteviol and the steviol glycosides, stevioside, rebaudioside A, rebaudioside C, and dulcoside A are responsible for inhibiting inflammation (Yasukawa et al., 2002; Mizushina et al., 2005).

Stevia's immunomodulatory activity, and anti-inflammatory properties were confirmed *in vitro* by the established decrease in TNF- α , IL-1 β , IL-6 synthesis and inhibiting of nuclear transcription factor-kappa B (NF- κ B), and *in vivo* by inhibiting NF- κ B and the mitogen-activated protein kinase (MAPK) signaling, and the release of proinflammatory cytokines (Fengyang et al., 2012; Jiang et al., 2019; Kasti et al., 2022).

The extracts used in a Korean-Indian experiment were established to provoke a non-specific immune response in rats, and furthermore improved humoral and cell-mediated immunity effectively (Shukla & Mehta, 2015). On that ground, the authors claim *S. rebaudiana* is a therapeuticly effective and safe immunomodulatory agent that can be used as a 'Rasayana' in traditional medicine. In adition, stevia is rich in valuable protein and amino acids, minerals and fibers (Abou-Arab et al., 2010; Wolwer-Rieck, 2012).

It has been demonstrated that stevia has inhibitory properties of its polyphenolic constituents (aglycones, steviol and isosteviol, and their metabolites) on tumor promotion and initiation by blocking Epstein-Barr virus early antigen (EBV-EA) induction and tumor formation (Konoshima & Takasaki, 2002; Akihisa et al., 2004; Takasaki et al., 2009). Stevioside, isosteviol, potently inhibit DNA replication and cancer cell growth in vitro (Mizushina et al., 2005). It is noteworthy that except non-carcinogenic, non-mutagenic effects, stevia glycosides are non-teratogenic and do not cause toxicity and genotoxicity (Momtazi-Borojeni et al., 2017; Sharif et al., 2017; EFSA FAF Panel, 2022). On regular basis, the European Food Safety Authority releases reports on stevia and its derivatives usage, based on scientific opinion and sets acceptable daily intake for steviol glycosides (E 960) of 4 mg per 1 kg of body weight per day expressed as steviol (EFSA FAF Panel, 2022).

The research team of Ramos-Tovar were the first to show that stevia can take control over Nrf2, thereby counteracting oxidative stress, and hence prevent necrosis and cholestasis, implying that it can be administered in the clinical treatment of acute and chronic liver diseases (Ramos-Tovar et al., 2018). Similarly, it was found that stevia has the potential to protect against oxidative stress and age-related conditions Pencho Penchev et al.

via the Akt/Nrf2/HO-1 pathway, as its residue extract significantly increased the activities of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and total antioxidant capacity (T-AOC), and decreased acetylcholinesterase (AChE) activity and malondialdehyde (MDA) level in mice (Zhao et al., 2019). The established clear membrane-stabilizing effect of stevia is to add to its antioxidant activity (Simonyan et al., 2021).

These many results of the effects of stevia as a medicinal plant imply that improvements in productivity of livestock should be also expected. The finding of its benefits on cholesterol and hence on cardiovascular healthcare suggest association between stevia-based diet and fatty acid profile of milk of dairy animals.

Stevia in monogastric animals

Studies in pigs (Munro et al., 2000; Geuns et al., 2003a; Wang et al., 2014) and poultry (Geuns et al., 2003b; Atteh et al., 2008) have demonstrated that stevioside supplementation leads to higher feed intake and utilization, as well as to positive effect on animal health. This can be associated with the findings that stevia and its glycosides can improve appetite in the porcine species (Wang et al., 2011, 2014; Guo et al., 2016; Yu et al., 2020), poultry (Jiang et al., 2020), as well as in horses (Ma & Ma, 2009).

Except in better consumption and utilization, some of these experiments resulted in improved health status of the digestive tract in pigs (Wang et al., 2014). It was found that 5% of stevia residue supplementation regulated poultry digestive function (Shang, 2011). Daily gain of 42-day broiler chickens supplemented 80 ppm of stevia-based sweetener was significantly higher (Molina-Barrios et al., 2021). In addition, immune response to Newcastle disease virus was also higher but not to infectious bronchitis.

A recent study of Liu et al. (2022) indicated that a supplementation doze of 200 mg/kg of stevia residue extract significantly reduced the incidence of diarrhea in piglets. A higher doze of 400 mg/kg significantly reduced serum MDA content and significantly increased the total antioxidant capacity, total superoxide dismutase, and glutathione peroxidase activity in the serum and catalase and glutathione peroxidase activity in the liver. Moreover, this higher doze improved health status by regulating antioxidant reduction homeostasis of the piglets, due to an increase of potentially beneficial bacteria like *Prevotellaceae* and *Roseburia*.

Similarly, a series of Bulgarian-British experiments showed that stevia could be effective against oxidative stress also in broilers. As supplemented to the diet, it improved liver antioxidant status via increasing liver vitamin E and carotenoids (Pirgozliev et al., 2021a) and via increasing dietary antioxidant availability (Pirgozliev et al., 2021b). In the third study (Pirgozliev et al., 2022), the addition of stevia to the diet of laying hens lead to a richer color of the yolks and to an increased content of carotenoids in the eggs, but did not affect the palatability of the stored eggs. The authors conclude that increasing the dietary sources of lutein and zeaxanthin for humans can be achieved by including a stevia supplement in feed for laying hens to increase the concentration of these macular carotenoids in egg yolks.

Studying stevia plant as a potential fodder in livestock, Atteh et al. (2011) found that the fat percentage in the leaves was similar to other non-oil plants (2.6%), and it had high degree of unsaturated fatty acids - significantly higher then in the stem. The protein content in the leaves (16%) was higher than in the stems (6.7%) – in both cases moderately digestible, so retained in the broilers' tract. The high crude fiber content and low metabolisable energy of the stems renders stevia a good supplement in low nutritional diets for monogastric animals, as well as a sweet energy diluent. In the ruminants, that can digest high fiber diets stevia stems can be used as a source of energy. This study shows that minerals were higher in stevia leaves than stems, except for of Na and K that are lower. Noteworthy is the high Fe content of the leaves that can be used in anemic conditions. So, the authors concluded that stevia protein and gross energy content might be suitable for ruminants rather than for monogastric animals like pigs and chickens.

Stevia in ruminants

The studies with *Stevia rebaudiana* B. as a medicinal plant on large and small ruminants are scarce.

The first known studies with stevia in ruminants are published in 2014. One of them shows the effect of a feed containing 6.5% of stevioside (as an essential oil) and 25% of yeast culture on rumen and rectal temperature in steers (Cho et al., 2014).

According to an Italian study, stevia extract has negative effect on the rumen protozoa population only at low dietary doses (Sarnataro & Spanghero, 2020).

Stevia is a medicinal plant that can be used as a natural feed with environmental and bioactive functions. In a study on sheep, He et al. (2017) describe the *in vitro* rumen fermentation of stevia residue material as typical acetic acid type fermentation, which is capable of promoting carbohydrate fermentation in the rumen, improving energy utilization and the production of volatile fatty acids. As a result, it affords lower methane emissions due to rumen fermentation. The authors also observe that stevia stimulates the microbe metabolism of the rumen, hence the production of microbial protein.

A recent trial on dairy cows in China showed the replacement of only 6% of alfalfa hay with stevia hay did not affect nutrients intake but improved the fiber digestibility, and hence increased milk yield, milk fat and ruminal volatile fatty acids, while the 12% stevia supplementation did not have such beneficial effects (Jiang et al., 2022).

Another Chinese experiment (Han et al., 2019) established that with the supplementation of 400 to 800 mg/kg stevioside to the diet of dairy goats, the appetite and feed intake increased as well as the digestibility of fiber and portion of VFAs in ruminal content. In terms of the effect on the production of ruminant origin, it was observed that in Korean native beef the inclusion of 1% (on weight basis) stevioside and selenium improved polyunsaturated fatty acid, lipid oxidation during the storage and microbial count; and also growth performance, carcass traits, and meat quality (Shin et al., 2021).

According to a very recent trial (Zhang et al., 2023), 1.0% stevia stalk supplementation improves feed conversion and promotes rumen fermentation, and another study (Xie & Wang, 2010) established that a supplementation of 0.3% to 0.6% whole stevia plants for one month could improve the appetite of dairy cows, especially important during the period of negative energy balance.

Conclusions

In this review we summarized the history of the medicinal applications of Stevia rebaudiana B. and its derivates in animals, and in livestock in particular and their effects on the products thereof. The objective of this review found few result in the available scientific literature with focus on stevia as a supplementation in ruminants, and the species studied are few as well. Still, these findings imply that stevia would presumably play similar role in other ruminants as well. Yet, this calls for new research in the field of antioxidant and fatty-acid properties of milk and meat as affected by stevia supplementation in some other species, not studied to date, like the buffalo. Here is the place to note that growing stevia in Bulgaria has its peculiarities. Its cultivation is vegetative and it is maintained under in vitro conditions and moreover it needs more efforts in the latitudes of Bulgaria, where it should be treated like an annual, not like perennial plant - involving autumn hibernation and spring implantation of roots, as well as further development within one season. Though, based on the knowledge at this point, it seems worth growing stevia for purposes like ruminant foodstuff production because of the many benefits thereof.

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References

- Abou-Arab, A. E., Abou-Arab, A. A. & Abu-Salem, M. F. (2010). Physico-chemical assessment of natural sweeteners steviosides produced from *Stevia rebaudiana* Bertoni plant. *African Jour*nal of Food Science, 4-5, 269–281.
- Aghajanyan, A., Movsisyan, Z. & Trchounian, A. (2017). Antihyperglycemic and antihyperlipidemic activity of hydroponic *Stevia rebaudiana* aqueous extract in hyperglycemia induced by immobilization stress in rabbits. *BioMed. Research International*, 2017(1), 9251358. DOI:10.1155/2017/9251358.
- Ahmad, N., Fazal, H., Abbasi, B. & Farooq, S. (2010). Efficient free radical scavenging activity of *Ginkgo biloba*, *Stevia rebaudiana* and *Parthenium hysterophorous* leaves through DPPH (2,2-diphe-nyl-1-picrylhydrazyl). *International Journal of Phytomedicine*, 2(3), 231–239.
- Ahmad, J., Khan, I., Blundell, R., Azzopardi, J. & Mahomoodally, M. F. (2020). Stevia rebaudiana Bertoni: an updated review of its health benefits, industrial applications and safety. *Trends in Food Science & Technology*, 100, 177–189.
- Akihisa, T., Hamasaki, Y., Tokuda, H., Ukiya, M., Kimura, Y. & Nishino, H. (2004). Microbial transformation of isosteviol and inhibitory effects on Epstein-Barr virus activation of the transformation products. *Journal of Natural Products*, 67(3), 407–410.
- Allam, S. M., El Hosseiny, H. M., Gawad, A. M. A., El-Saadany, S. A. & Zeid, A. M. M. (1999). Medicinal herbs and plants as feed additives for ruminant. I. Effect of using some medicinal herbs and plants as feed additives on Zaraibi goat performance. *Egyptian Journal of Nutrition and Feeds, 2*(Special Issue), 349–365.
- Ashes, J. R., Welch, P. S. V., Gulati, S. K., Scott, T. W. & Brown, G. H. (1992). Manipulation of the fatty acid composition of milk by feeding protected canola seeds. *Journal of Daily Science*, 75(4), 1090–1096.
- Atteh, J. O., Onagbesan, O. M., Tona, K., Decuypere, E., Geuns, J. M. & Buyse, J. (2008). Evaluation of supplementary stevia (*Stevia rebaudiana* Bertoni) leaves and stevioside in broiler diets: effects on feed intake, nutrient metabolism, blood parameters and growth performance. *Journal of Animal Physiology* and Animal Nutrition, 92(6), 640–649.
- Atteh, J. O., Onagbesan, O. M., Tona, K., Buyse, J., Decuypere, E. & Geuns, J. M. (2011). Potential use of *Stevia rebaudiana* in animal feeds. *Archivos de Zootecnia*, 60(229), 133–136.
- Bauman, D. E., Mather, I. H., Wall, R. J. & Lock, A. L. (2006). Major advances associated with the biosynthesis of milk. *Jour-*

nal of Dairy Science, 89(4), 1235-1243.

- Belury, M. A. (2002). Dietary conjugated linoleic acid in health: Physiological effects and mechanisms of action. *Annual Review* of Nutrition, 22(1), 505–531.
- Bender, C., Graziano, S. & Zimmermann, B. F. (2015). Study of *Stevia rebaudiana* Bertoni antioxidant activities and cellular properties. *International Journal of Food Sciences and Nutrition*, 66(5), 553–558.
- Benford, D. J., DiNovi, M. & Schlatter, J. (2006). Safety evaluation of certain food additives: Steviol Glycosides" (PDF – 18MB). WHO Food Additives Series (World Health Organization Joint FAO/WHO Expert Committee on Food Additives (JECFA) 54.
- Beyero, N., Kapoor, V. & Tewatia, B. S. (2015). Effect of different roughage: Concentrate ratio on milk yield and its fatty acid profile in dairy cows. *Journal of Biology, Agriculture and Healthcare, 5*(13), 176–185.
- **Brandie, J.** (2004). FAQ Stevia, nature, s natural low calorie sweetener. *Agriculture and Agri-Food*, Canada. http://res2.agr. ca/London/faq/ stevia e.htm.
- Carakostas, M., Curry, L., Boileau, A. & Brusick, D. (2008). Overview: The history, technical function and safety of rebaudioside A, a naturally occurring steviol glycoside, for use in food and beverages. *Food and Chemical Toxicology*, 46(7), S1– S10 DOI: 10.1016/j.fct.2008.05.003.
- Cedillo, J., Kholif, A. E., Salem, A. Z. M., Elghandour, M. M. Y., Vázquez, J. F., Alonso, M. U., Barbabosa, A., Chagoyán, J. C. V. & Reyna, A. G. (2015). Oral administration of Sauce llorón extract to growing lambs to control gastrointestinal nematodes and Moniezia spp. Asian Pacific Journal of Tropical Medicine, 8(7), 520–525.
- Chatsudthipong, V. & Muanprasat, C. (2009). Stevioside and related compounds: Therapeutic benefits beyond sweetness. *Pharmacology & Therapeutics*, 121(1), 41–54.
- Cho, B. O., Ryu, H. W., So, Y., Cho, J. K., Woo, H. S., Jin, C. H., Seo, K. I., Park, J. C. & Jeong, I. Y. (2013). Anti-inflammatory effect of austroinulin and 6-O-acetyl-austroinulin from *Stevia rebaudiana* in lipopolysaccharide-stimulated RAW2647 macrophages. *Food and Chemical Toxicology*, 62, 638–644.
- Cho, S., Mbiriri, D. T., Shim, K., Lee, A. L., Oh, S. J., Yang, J., Ryu, C., Kim, Y. H., Seo, K. S., Chae, J. I., Oh, Y. K. & Choi, N. J. (2014). The influence of feed energy density and a formulated additive on rumen and rectal temperature in Hanwoo steers. *Asian-Australasian Journal of Animal Sciences*, 27(11), 1652–1662.
- Chu, Y., Chang, C. & Hsu, H. (2000). Flavonoid content of several vegetables and their antioxidant activity. *Journal of the Science of Food and Agriculture*, 80(5), 561–566.
- Curi, R., Alvarez, M., Bazotte, R. B., Botion, L. M., Godoy, J. L. & Bracht, A. (1986). Effect of *Stevia rebaudiana* on glucose tolerance in normal adult humans. *Brazilian Journal of Medical* and *Biological Research*, 19(6), 771–774.
- Daley, C. A., Abbott, A., Doyle, P. S., Nader, G. A. & Larson, S. (2010). A review of fatty acid profiles and antioxidant content in grass-fed and grain-fed beef. *Nutrition Journal*, 9, 1-12. https://doi.org/10.1186/1475-2891-9-10
- Diasio, R. B. & LoBuglio, A. F. (1996). Immunomodulator: immu-

nosuppressive agents and I immunostimulants. *In:* Goodman & Gilman's, The Pharmacological Basis of Therapeutics, ed. by Hardman, J.G. and Limbird, L.E. The New York: *McGraw-Hill*, 9th Edition, 1291–1307.

- Edeoga, H. O., Okwu, D. E. & Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*, 4(7), 685–688.
- **EFSA FAF Panel** (EFSA Panel on Food Additives and Flavourings) (2022). Safety evaluation of glucosylated steviol glycosides as a food additive in different food categories. EFSA Journal, Wiley-VCH GmbH, 20(2), 7066.
- Enchev, S., Mehmed, A. & Kikidonov, G. (2018). Effect of mineral and organic fertilization on the production of Stevia (*Stevia rebauduana* B.). *Bulg. J. Agric. Sci., 24* (Suppl. 2), 100–103.
- Fahim, N. H., Kholif, A. E. & Azzaz, H. H. (2022). Fennel and ginger improved nutrient digestibility and milk yield and quality in early lactating Egyptian buffaloes. *Annals of Animal Science*, 22(1), 255–270.
- FAO/WHO (Food and Agriculture Organization/World Health Organization) (2003). Diet, nutrition and prevention of chronic diseases. WHO Technical Report Series #916, Report of a joint WHO/FAO expert conclusion, Geneva 2003, 149.
- Fengyang, L., Yunhe, F., Bo, L., Zhicheng, L., Depeng, L., Dejie, L., Wen, Z., Yongguo, C., Naisheng, Z., Xichen, Z. & Zhengtao, Y. (2012). Stevioside suppressed inflammatory cytokine secretion by downregulation of NF-κB and MAPK signaling pathways in LPS-stimulated RAW264.7 cells. *Inflammation*, 35, 1669–1675.
- Fernandes, S. A. A., Mattos, W. R. S., Matarazzo, S. V., Tonhati, H., Gama, M. A. S. & Lanna, D. P. D. (2007). Total fatty acids in Murrah buffaloes milk on commercial farms in Brazil. *Italian Journal of Animal Science*, 6(Suppl. 2), 1063–1066.
- Ferri, L. A., Alves-Do-Prado, W., Yamada, S. S., Gazola, S., Batista, M. R. & Bazotte, R. B. (2006). Investigation of the antihypertensive effect of oral crude stevioside in patients with mild essential hypertension. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 20(9), 732–736.
- Geuns, J. M., Augustijns, P., Mols, R., Buyse, J. G. & Driessen,
 B. (2003a). Metabolism of stevioside in pigs and intestinal absorption characteristics of stevioside, rebaudioside A and steviol. *Food Chemistry and Toxicology*, 41(11), 1599–1607.
- Geuns, J. M., Malheiros, R. D., Moraes, V. M., Decuypere, E. M., Compernolle, F. & Buyse, J. G. (2003b). Metabolism of stevioside by chickens. *Journal of Agricultural and Food Chemistry*, 51(4),1095–1101.
- Ghanta, S., Banerjee, A., Poddar, A. & Chattopadhyay, S. (2007). Oxidative DNA damage preventive activity and antioxidant potential of *Stevia rebaudiani* (Bertoni), a natural sweetener. *Journal of Agricultural and Food Chemistry*, 55(26), 10962–10967.
- Ghosh, S., Subudhi, E. & Nayak, S. (2008). Antimicrobial assay of *Stevia rebaudiana* Bertoni leaf extracts against 10 pathogens. *International Journal of Integrative Biology*, 2(1), 27–31.
- Givens, D. I. & Shingfield, K. J. (2006). Optimizing dairy milk fatty acid composition. *In:* Improving the Fat Content of Foods, ed. by Williams, C. and Buttriss, J., *Woodhead Publishing Lim-*

ited. Cambridge, 252-280.

- Goyal, S. K., Samsher, G. R. & Goyal, R. K. (2010). Stevia (Stevia rebaudiana) a bio-sweetener: a review. International Journal of Food Sciences and Nutrition, 61(1), 1–10.
- Gregersen, S., Jeppesen, P. B., Holst, J. J. & Hermansen, K. (2004). Antihyperglycemic effects of stevioside in type 2 diabetic subjects. *Metabolism: Clinical and Experimental*, 53(1), 73–76.
- Guo, L. R., Su, Z., Kong, Z. W., Huang, J. F., Zhong, R. Y. & Guo, T. F. (2016). Effects of feeding fermented Stevia residue on amino acid and fatty acid composition in pig muscle. *Pig Science*, 33, 72-73.
- Gupta, E., Purwar, S., Sundaram, S., Tripathi, P. & Rai, G. (2016). Stevioside and rebaudioside a - predominant entkaurene diterpene glycosides of therapeutic potential: a review. Czech Journal of Food Science, 34(4), 281–299.
- Han, X., Chen, C., Zhang, X., Wei, Y., Tang, S., Wang, J., Tan, Z. & Xu, L. (2019). Effects of dietary stevioside supplementation on feed intake, digestion, ruminal fermentation, and blood metabolites of goats. *Animals*, 9(2), 32.
- He, L., Ji, S. S., Zhang, Z. H., Wang, D. S., Guo, L. R., Zhang, G. H. & Ding, J. N. (2017). Effects of Andrographis paniculata residue and sweet leaf inulin residue on sheep rumen fermentation in vitro. Chinese Journal of Animal Husbandry, 53, 86–89.
- Ilias, N., Hamzah, H., Ismail, I. S., Mohidin, T. B. M., Idris, F. & Ajat, M. (2021). An insight on the future therapeutic application potential of *Stevia rebaudiana* Bertoni for atherosclerosis and cardiovascular diseases. *Biomedicine & Pharmacotherapy*, 143. DOI: 10.1016/j.biopha.2021.112207.
- Ilieva, Y., Mihaylova, D., Ilyazova, A. & Penchev, P., Abadjieva, D. & Kistanova, E. (2022). Effects of the herbal preparation AyuFertin, used for anestrus overcome, on fatty acids composition of milk in Bulgarian Murrah buffaloes. *Bulgarian Journal* of Veterinary Medicine, 25(3), 440-450.
- Jiang, J., Qi, L., Lv, Z., Jin, S., Wei, X. & Shi, F. (2019). Dietary stevioside supplementation alleviates lipopolysaccharide-induced intestinal mucosal damage through anti-inflammatory and antioxidant effects in broiler chickens. *Antioxidants*, 8(12), 575.
- Jiang, J. L., Qi, L. N., Dai, H. J., Hu, C. H, Lu, Z. P., Wei, Q. W. & Shi, F. X. (2020). Dietary stevioside supplementation improves laying performance and eggshell quality through increasing estrogen synthesis, calcium level and antioxidant capacity of reproductive organs in aged breeder hens. *Animal Feed Science and Technology*, 269, 114682.
- Jiang, M., Datsomor, O., Cheng, Z., Meng, Z., Zhan, K., Yang, T., Huang, Y., Yan, Q. & Zhao, G. (2022). Partial substitution of alfalfa hay by Stevia (*Stevia rebaudiana*) hay can improve lactation performance, rumen fermentation, and nitrogen utilization of dairy cows. *Frontiers in Veterinary Science*, 9, 899148. DOI: 10.3389/fvets.2022.899148.
- Kalač, P. & Samková, E. (2010). The effects of feeding various forages on fatty acid composition of bovine milk fat: a review. *Czech Journal of Food Science*, 55(12), 521–537.
- Kasti, A. N., Nikolaki, M. D., Synodinou, K. D., Katsas, K. N., Petsis, K., Lambrinou, S., Pyrousis, I. A. & Triantafyllou,

K. (2022). The effects of stevia consumption on gut bacteria: friend or foe? *Microorganisms*, *10*(4), 744. DOI: 10.3390/microorganisms10040744.

- Khattab, M. S. A., Kholif, A. E., Abd El Tawab, A. M., Shaaban,
 M. M., Hadhoud, F. I., El-Fouly, H. A. & Olafadehan, O.
 A. (2020). Effect of replacement of antibiotics with thyme and celery seed mixture on the feed intake and digestion, ruminal fermentation, blood chemistry, and milk lactation of lactating Barki ewes. *Food & Function*, 11(8), 6889–6898.
- Kikindonov, Tz. (2013). Assessment of initial material for stevia (Stevia rebaudiana B.) breeding. Agricultural Science and Technology, 5(1), 22–24, 1313-8820.
- Kikindonov, Tz. & Enchev, S. (2012). Assessment of seeds germination of stevia (*Stevia rebaudiana* B.) origins. *Agricultural Science*, 45(3), 18–23 (Bg).
- Konoshima, T. & Takasaki, M. (2002). Cancer-chemopreventative effects of natural sweeteners and related compounds. *Pure* and Applied Chemistry, 74(7), 1309-1316.
- Kurek, J. M., Mikolajczyk-Stecyna, J. & Krejpcio, Z. (2023). Steviol glycosides from *Stevia rebaudiana* Bertoni mitigate lipid metabolism abnormalities in diabetes by modulating selected gene expression – An *in vivo* study. *Biomedicine & Pharmacotherapy*, 166, 115424 DOI:10.1016/j.biopha.2023.115424.
- Lemus-Mondaca, R., Vega-Galvez, A., Zura-Bravo, L. & Ah-Hen, K. (2012). *Stevia rebaudiana* Bertoni, source of a high-potency natural sweetener: a comprehensive review on the biochemical, nutritional and functional aspects. *Food Chemistry*, 132(3), 1121–1132.
- Lemus-Mondaca, R., Vega-Galvez, A., Rojas, P., Stucken, K., Delporte, C., Valenzuela-Barra, G., Jagus, R. J., Agüer, M. V. & Pasten, A. (2018). Antioxidant, antimicrobial and anti-inflammatory potential of *Stevia rebaudiana* leaves: effect of different drying methods. *Journal of Applied Research on Medicinal and Aromatic Plants, 11*, 37–46.
- Liu, S., Xiong, Y., Cao, S., Wen, X., Xiao, H., Li, Y., Chi, L., He, D., Jiang, Z. & Wang, L. (2022). Dietary stevia residue extract supplementation improves antioxidant capacity and intestinal microbial composition of weaned piglets. *Antioxidants*, 11(10), 2016. DOI: 10.3390/antiox11102016.
- Ma, L. & Ma, Y. (2009). The comprehensive development and utilization of stevia. *China Sugar*, 1, 68–69.
- Matloup, O. H., Abd El Tawab, A. M., Hassan, A. A., Hadhoud, F. I., Khattab, M. S. A., Khalel, M. S., Sallam, S. M. A. & Kholif, A. E. (2017). Performance of lactating Friesian cows fed a diet supplemented with coriander oil: Feed intake, nutrient digestibility, ruminal fermentation, blood chemistry, and milk production. *Animal Feed Science & Technology*, 226, 88–97.
- Mehmed, A. & Enchev, S. (2020). Influence of organic foliar fertilizerson stevia development (*Stevia rebaudiana* B.). *Journal* of Mountain Agriculture on the Balkans, 23(4), 230-242.
- Mehmed, A., Enchev, S. & Bozhimirov, S. (2021). Assessment of initial material for stevia selection (*Stevia rebaudiana* B.). *Bulg. J. Agric. Sci.*, 27(3), 536–540.
- Midmore, D. J. & Rank, A. H. (eds). (2002). A new rural industry – Stevia – to replace imported chemical sweeteners. A Report for the Rural Industries Research and Development Corporation. Barton, Queensland, Australia.

- Mizushina, Y., Akihisa, T., Ukiya, M., Hamasaki, Y., Murakami-Nakai, C., Kuriyama, I., Takeuchi, T., Sugawara, F. & Yoshida, H. (2005). Structural analysis of isosteviol and related compounds as DNA polymerase and DNA topoisomerase inhibitors. *Life Sciences*, 77(17), 2127–2140.
- Molina-Barrios, R. M., Avilés-Trejo, C. R., Puentes-Mercado, M. E., Cedillo-Cobián, J. R. & Hernández-Chavez, J. F. (2021). Effect of dietary stevia-based sweetener on body weight and humoral immune response of broiler chickens. *Veterinary World*, 14(4), 913–917.
- Momtazi-Borojeni, A. A., Esmaeili, S., Abdollahi, E. & Sahebkar, A. (2017). A review on the pharmacology and toxicology of steviol glycosides extracted from *Stevia rebaudiana*. *Current Pharmaceutical Design*, 23(11), 1616–1622.
- Muanda, F. N., Soulimani, R., Diop, B. & Dicko, A. (2011). Study on chemical composition and biological activities of essential oil and extracts from *Stevia rebaudiana* Bertoni leaves. *LWT* -Food Science and Technology, 44(9), 1865–1872.
- Munro, P. J., Lirette, A., Anderson, D. M. & Ju, H. Y. (2000). Effects of a new sweetener, Stevia, on performance of newly weaned pips. *Canadian Journal of Animal Science*, 80(3), 529–531.
- Nikolova-Damyanova, B., Bankova, V. & Popov, S. (1994). Separation and quantitation of stevioside and rebaudiside A in plant extracts by normal-phase high performance liquid chromatography and thin-layer chromatography: A comparison. *Phytochemical Analysis*, 5(2), 81–85.
- Nogoy, K. M. C., Sun, B., Shin, S., Lee, Y., Zi Li, X., Choi, S. H. & Park, S. (2022). Fatty acid composition of grain- and grass-fed beef and their nutritional value and health implication. *Food Science of Animal Resources*, 42(1), 18-33.
- Park, J. E. & Cha, Y. S. (2010). Stevia rebaudiana Bertoni extract supplementation improves lipid and carnitine profiles in C57BL/6J mice fed a highfat diet. J. Sci. Food Agric., 90(7), 1099–1105.
- Parodi, P. W. (2004). Milk fat in human nutrition. *Australian Journal of Dairy Technology*, 59(1), 3.
- Patwardhan, B., Kalbag, D., Patki, P. S., Nagsampagi, B. A. (1990). Search of immunomodulatory agents: a review. *Indian* Drugs, 28(2), 56-63.
- Penchev, P., Ilieva, Y. & Ivanova, S. (2022). Effect of supplementation of curcumin to the diet of buffaloes on the fatty-acid profile of milk and the derivative yoghurt. *Zhivotnovadni nauki*, 59(4), 78–87 (Bg).
- Pirgozliev, V., Whiting, I. M., Mansbridge, S. C., Enchev, S., Rose, S. P., Kljak, K., Johnson, A. E., Drijfhout, F., Orczewska-Dudek, S. & Atanasov, A. G. (2021a). Effect of rearing temperature on physiological measures and antioxidant status of broiler chickens fed stevia (*Stevia rebaudiana* B.) leaf meal and exogenous xylanase. *Current Research in Biotechnology*, 3, 173-181.
- Pirgozliev, V., Kljak, K., Whiting, I. M., Rose, S. P., Mansbridge, S. C., Enchev, S., Atanasov, A. G. & Stringhini, H. (2021b). Feeding dry stevia leaf (*Stevia rebaudiana*) or xylanase improves the hepatic antioxidative status of broiler chickens. *Research in Veterinary Science*, 136, 227–229.
- Pirgozliev, V., Whiting, I. M., Kljak, K., Mansbridge, S. C., Ata-

nasov, A. G., Rose, S. P. & Enchev, S. (2022). Stevia (*Stevia rebaudiana*) improves carotenoid content in eggs when fed to laying hens. *Foods*, *11*, 1418.

- Puri, M., Sharma, D. & Tiwari, A. K. (2011). Downstream processing of stevioside and its potential applications. *Biotechnol*ogy Advances, 29(6), 781–791.
- Ramos-Tovar, E., Hernández-Aquino, E., Casas-Grajales, S., Buendia-Montaño, L. D., Galindo-Gómez, S., Camacho, J., Tsutsumi, V. & Muriel, P. (2018). Stevia prevents acute and chronic liver injury induced by carbon tetrachloride by blocking oxidative stress through Nrf2 upregulation. Oxidative Medicine and Cellular Longevity, 2018(1), 3823426. DOI: 10.1155/2018/3823426.
- Rotimi, S. O., Rotimi, O. A., Adelani, I. B., Onuzulu, C., Obi, P. & Okungbaye, R. (2018). Stevioside modulates oxidative damage in the liver and kidney of high fat / low streptozocin diabetic rats. *Heliyon*, 4(5), e00640. DOI: 10.1016/j.heliyon. 2018.e00640
- Salem, A. Z. M., Elghandour, M. M. Y., Kholif, A. E., López, S., Pliego, A. B., Cipriano-Salazar, M., Chagoyán, J. C. V., de Oca Jiménez, R. M. & Alonso, M. U. (2017). Tree leaves of Salix babylonica extract as a natural anthelmintic for small-ruminant farms in a semiarid region in Mexico. Agroforestry Systems, 91, 111–122.
- Sarnataro, C. & Spanghero, M. (2020). In vitro rumen fermentation of feed substrates added with chestnut tannins or an extract from Stevia rebaudiana Bertoni. Animal Nutrition, 6(1), 54–60.
- Savita, S. M., Sheela, K., Sunanda, S., Shankar, A. G., Ramakrishna, P. & Sakey, S. (2004). Health implications of Stevia rebaudiana. Journal of Human Ecology, 15(3), 191–194.
- Scollan, N. D., Enser, M., Gulati, S.K., Richardson, I. & Wood, J. D. (2003). Effects of including a ruminally protected lipid supplement in the diet on the fatty acid composition of beef muscle. *British Journal of Nutrition*, 90(3), 709–716.
- Sehar, I., Kaul, A., Bani, S., Pal, H. C. & Saxena, A. K. (2008). Immune up regulatory response of a non-caloric natural sweetener, stevioside. *Chemico-Biological Interactions*, 173(2), 115–121.
- Shang, H. Q. (2011). Research progress on comprehensive utilization of Stevia. *Biology Teaching*, 36, 4–6.
- Sharif, R., Chan, K. M., Ooi, T. C. & Mohammad, N. F. (2017). Cytotoxicity and genotoxicity evaluation of stevioside on CCD18Co and HCT 116 cell lines. *International Food Research J.*, 24(1), 341–345.
- Sharma, N., Mogra, R. & Upadhyay, B. (2009). Effect of stevia extract intervention on lipid profile. *Studies on Ethno-Medicine*, 3(2), 137–140.
- Shin, Y. G., Rathnayake, D., Mun, H. S., Dilawar, M. A., Pov, S. & Yang, C. J. (2021). Sensory attributes, microbial activity, fatty acid composition and meat quality traits of Hanwoo cattle fed a diet supplemented with stevioside and organic selenium. *Foods*, 10(1), 129. DOI: 10.3390/foods10010129.
- Shingfield, K. J., Salo-Väänänen, P., Pahkala, E., Toivonen, V., Jaakkola, S., Piironen, V. & Huhtanen, P. (2005). Effect of forage conservation method, concentrate level and propylene glycol on the fatty acid composition and vitamin content of cows'mik. *Journal of Dairy Research*, 72(3), 349–361.

- Shivanna, N., Naika, M., Khanum, F. & Kaul, V. K. (2013). Antioxidant, anti-diabetic and renal protective properties of *Stevia rebaudiana*. *Journal of Diabetes and its Complications*, 27(2), 103–113.
- Shukla, S. & Mehta, A. (2015). Comparative phytochemical analysis and *in vivo* immunomodulatory activity of various extracts of *Stevia rebaudiana* leaves in experimental animal model. *Frontiers in Life Science*, 8, 55-63.
- Shukla, S., Mehta, A., Mehta, P. & Bajpai, V. K. (2012). Antioxidant ability and total phenolic content of aqueous leaf extract of *Stevia rebaudiana* Bertoni. *Experimental and Toxicologic Pathology*, 64(7-8), 807–811.
- Simonyan, K. V., Chavushyan, V. A., Avetisyan, L. G., Simonyan, G. M., Hovhannisyan, L. E. & Simonyan, M. A. (2021). Regulatory effects of *Stevia rebaudiana* on NADPH oxidase-related manifestations of oxidative stress in diabetic rats with spinal cord injury. *Neurophysiology*, 53(1), 13–21.
- Singh, S. & Rao, G. (2005). Stevia: The herbal sugar of 21st Century. Sugar Technology, 71(1),17–24.
- Slavova, J., Nenkova, D. & Ivanova, I. (2003). Study on the benzymidazol upon Stevia rebaudiana Bertoni, cultivated in vitro. Bulg. J. Agric. Sci., 9(2), 225–228.
- Smith, S. B., Clare, A. G., Lunt, D. K. & Brooks, M. A. (2009). Regulation of fat and fatty acid composition in beef cattle. *Asian-Australian Journal of Animal Science*, 22(9), 1225– 1233.
- Takahashi, K., Matsuda, M., Ohashi, K., Taniguchi, K., Nakagomi, O., Abe, Y., Mori, S., Sato, N., Okutani, K. & Shigeta, S. (2001). Analysis of anti-rotavirus activity of extract from *Stevia rebaudiana*. *Antiviral Research*, 49(1), 15–24.
- Takasaki, M., Konoshima, T., Kozuka, M., Tokunda, H. & Takayasu, J. (2009). Cancer preventive agents. Part 8: Chemopreventive effects of stevioside and related compounds. *Bioor*ganic & Medicinal Chemistry, 17(2), 600–605.
- Tanova, K. & Kaschieva, M. (2018). Testing alternative control means for the pathogenic fungus *Alternaria alternate* f. ssp. stevae, isolated from stevia – Stevia rebaudiana Bertoni. Journal of Mountain Agriculture on the Balkans, 21(6), 214–223.
- Thomas, J. & Glade, M. (2010). Stevia: It's not just about calories. Open Obesity Journal, 2, 101–109.
- Tomita, T., Sato, N., Arai, T., Shiraishi, H., Sato, M., Takeuchi, M. & Kamio, Y. (1997). Bactericidal activity of a fermented hot-water extract from *Stevia rebaudiana* Bertoni towards enterohemorrhagic *Escherichia coli* O157:H7 and other foodborne pathogenic bacteria. *Microbiology and Immunology*, 41(12), 1005–1009.
- Uchkunov, V., Uchkunova, K. & Uchkunov, I. (2015). Effect of the selection on the degree of resistance to *Alternaria alternata* f. ssp. *steviae* in breeding origins of stevia (*Stevia rebaudiana* B.). *Plant Science*, 52(1), 36–38 (Bg).
- Wang, Y. X., Zhou, G. L., Liao, Z. Y., Xu, W., Dong, Q. G. & Wang, T. (2011). Study on the application of stevioside in weaning piglets. *China Feed*, 4, 29-31.
- Wang, L. S., Shi, Z., Shi, B. M. & Shan, A. S. (2014). Effects of dietary stevioside/rebaudioside A on the growth performance and diarrhea incidence of weaned piglets. *Animal Feed Science* & *Technology*, 187, 104–109.

- Wolwer-Rieck, U. (2012). The leaves of *Stevia rebaudiana* (Bertoni), their constituents and the analyses thereof: a review. *Journal of Agricultural and Food Chemistry*, 60(4), 886–895.
- Xi, Y., Yamaguchi, T., Sato, M. & Takeuchi, M. (1998a). Antioxidant activity of Stevia rebaudiana. The Japanese Society for Food Science and Technology, 45(5), 310-316.
- Xi, Y., Yamaguchi, T., Sato, M. & Takeuchi, M. (1998b). Antioxidant mechanism of *Stevia rebaudiana* extract and antioxidant activity of inorganic salts. *The Japanese Society for Food Science and Technology*, 45(5), 317–322.
- Xie, F. Y. & Wang, G. J. (2010). Effects of adding stevia stalk in dairy cow diets on lactation performance of dairy cows. *Fuji*an Journal of Animal Husbandry and Veterinary Medicine, 12, 106.
- Yasukawa, K., Kitanaka, S. & Seo, S. (2002). Inhibitory effect of stevioside on tumor promotion by 12-O-tetradecanoylphor-

bol-13-acetate in two stage carcinogenesis in mouse skin. *Biological and Pharmaceutical Bulletin*, 25(11), 1488-1490.

- Yu, M., Tie, G., Liu, Z. & Diao, X. P. (2020). Effects of dietary supplementation with high fiber (*Stevia residue*) on the fecal flora of pregnant sows. *Animals*, 10(12), 2247 DOI: 10.3390/ ani10122247.
- Zhang, X., Jiao, T., Ma, S., Chen, X., Wang, Z., Zhao, S. & Ren, Y. (2023). Effects of different proportions of stevia stalk on nutrient utilization and rumen fermentation in ruminal fluid derived from sheep. *Peer Journal*, *11*. e14689 DOI: 10.7717/ peerj.14689.
- Zhao, L., Yang, H., Xu, M., Wang, X., Wang, C., Lian, Y., Mehmood, A. & Dai, H. (2019). Stevia residue extract ameliorates oxidative stress in d-galactose-induced aging mice via Akt/Nrf2/HO-1 pathway. *Journal of Functional Foods*, 52, 587–595.

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