

## **IMPROVEMENT OF PRODUCTIVE CAPACITY OF SOLONETZ BY APPLICATION OF MICROORGANISMS**

T. HAJNAL-JAFARI, S. DURIC, D. STAMENOV and M. MANOJLOVIC  
*University of Novi Sad, Faculty of Agriculture, 21000 Novi Sad, Serbia*

### **Abstract**

HAJNAL-JAFARI, T., S. DURIC, D. STAMENOV and M. MANOJLOVIC, 2015. Improvement of productive capacity of solonetz by application of microorganisms. *Bulg. J. Agric. Sci.*, 21: 767–770

Solonetz are dark alkaline soils which are mainly bare or covered by grasses. These sodic soils are characterized by pore water regime, high sodium saturation that is harmful to plants, unfavorable chemical and physical condition which makes them unsuitable for agricultural uses. Productive capacity of solonetz soils are usually improved by different meliorative measures such as incorporation of gypsum, deep ploughing or planting of a sodium-resistant crop. Application of microbial inocula could be a possible solution for soil quality improvement. The microbiological properties of solonetz from Kumane region were investigated as well as the effectiveness of four isolates of *Azotobacter* sp. and two isolates of *Bacillus* sp. Bacteria were isolated from virgin solonetz soil which was not covered by plants and was not recultivated. Pure cultures of each isolate were obtained containing  $10^{10}$  CFU/ml of *Azotobacter* sp. and  $10^{12}$  CFU/ml of *Bacillus* sp., respectively. Soil samples were inoculated. The virgin solonetz soil without inoculation was the control treatment. Soil sampling was performed 30 days after inoculation. Microbiological analyses included the determination of number and enzymatic activity of microorganisms. All six investigated bacteria strains stimulated the microbiological activity of solonetz. The total number of bacteria, number of fungi, actinomycetes, free-living nitrogen fixing bacteria and aminoheterotrophs were increased in relation to the control. Dehydrogenase activity was also higher after bacterial inoculation. This research highlighted the positive effect of *Azotobacter* sp. and *Bacillus* sp. application. Further research needs to be carried out in order to increase the benefits of microbial inoculation in semi controlled and field conditions.

*Key words:* solonetz, bacteria, inoculation, microbiological activity

### **Introduction**

Solonetz are soils with a dense, strongly structured, clay illuviation horizon that has a high proportion of adsorbed sodium and/or magnesium ions. ABtnC- and AEBtnC-profiles with a black or brown surface soil over a nitric horizon that starts at less than 100 cm from the soil surface. A calcic or gypsic horizon may be present below the natric horizon. Many solonetz have a field-pH around 8.5 indicative of the presence of free sodium carbonate. High levels of exchangeable sodium ions affect arable cropping, either directly (Na-toxicity) or indirectly, e.g. because of structure deterioration when soil material with a high proportion of adsorbed sodium and/or magnesium ions is wetted. Many solonetz in temperate regions have a humus-rich surface soil and can be used for arable farming or grazing. Solonetz is a soil type which occu-

pies significant area in Vojvodina province (80.33 ha, approx. 3.75%) (Sekulić et al., 2005). These are alkaline soils mostly used as natural pasture.

The suitability of 'virgin' solonetz for agricultural uses is almost entirely dictated by the depth and properties of the surface soil (FAO report, 2006). A 'deep' (say > 25 cm) humus-rich surface soil is needed for successful arable crop production. Unfortunately, most solonetz have only a much shallower surface horizon, or have lost the surface horizon altogether. By applying different meliorative measures the productive capacity of solonetz can be improved and used as cultivated soil (Belić, 1999; Belić et al., 2003). Solonetz amelioration has two basic elements such as improvement of the porosity of the (sub) surface soil, and lowering of the 'Exchangeable Sodium Percentage' ESP. Most reclamation attempts start with incorporation of gypsum or, exception-

ally, calcium chloride in the soil (Kruzhillin and Kazakova, 2011). Traditional reclamation strategies start with the planting of a sodium-resistant crop, e.g. *Rhodes* grass, to gradually improve the permeability of the soil.

Application of microbial inocula could be a possible solution for soil quality improvement. Since halomorph soils are present in Vojvodina, aside traditional amelioration, the application of microorganisms could be a potentially good measure for enhancing soil productive capacity. In this work, the effectiveness of several indigenous bacteria isolates (*Azotobacter* and *Bacillus*) was investigated.

## Materials and Methods

Soil samples were taken from Kumane region (experimental field Kumane-Faculty of Agriculture) with sterile soil sampler at the depth of 10, 20 and 30 cm. The samples were put in sterile polyethylene bags and transported to the Microbiology Laboratory. The number of microorganisms was determined by agar plate method, inoculating selective media with soil suspension of appropriate dilution (Trolldenier, 1996). The number of *Azotobacter* (AZB), number of aminoheterotrophs (AMH), total number of bacteria (TNB), number of actinomycetes (ACT) and number of fungi (FNG) were analyzed. The dehydrogenase activity (DHA) was determined spectrophotometrically by method of Thalmann (1968) modified by Lehnard (1956). Bacteria (*Azotobacter* and *Bacillus*) were isolated from virgin solonetz soil which was not covered by

plants and was not recultivated. Pure cultures of each isolates were obtained on Fjodorov medium and MPB medium containing  $10^{10}$  CFU of *Azotobacter* sp. and  $10^{12}$  CFU of *Bacillus* sp., respectively. Soil samples were inoculated. The virgin solonetz soil without inoculation was the control treatment. Soil sampling was performed 30 days after inoculation. Microbiological analyses included the determination of number and enzymatic activity of microorganisms.

## Results

Bulk and rhizosphere soil, as well surface and subsurface soil profiles, differ in their chemical, physical as well as biological properties (pH, salinity, redox potential, porosity, water retention, abundance and diversity of microbial species, etc) (Table 1).

Productive capacity of solonetz soils are usually improved by different meliorative measures such are incorporation of gypsum, deep plowing or planting of a sodium-resistant crop. Application of microbial inocula is a possible solution for soil quality improvement.

Microbiological activity of virgin solonetz is presented in Table 2. The top soil is characterizes by high total number of bacteria. Moreover, aminoheterotrophs are dominant in soil. Fungi and actinomycetes as major organic matter decomposers are also present in substantial number. *Azotobacter* is a free living nitrogen fixing bacteria and are present in soil. Since bacteria from N cycle were dominant in soil, several *Azoto-*

**Table 1**  
Physical and chemical properties of solonetz

Horizon	Depth, cm	pH		CaCO <sub>3</sub> , %	Humus, %	P <sub>2</sub> O <sub>5</sub> , mg/100g	K <sub>2</sub> O,	Ecc 25°C, ms/cm	Salts, %	pH, paste
		H <sub>2</sub> O	1MKCl							
Aoh/E,na	0-13	5.9	4.6	0	6.05	7.3	87.6	0.71	0.03	5.57
	13-36	7.86	6.38	0.27	1.69	1.5	17.6	2.69	0.14	7.07
	36-58	8.2	6.87	0	1.48	5.4	23.4	3.6	0.19	7.69
	58-85	8.93	7.3	0.42	0.83	6.4	28	2.15	0.17	8.21
Bt, na	85-114	9.31	7.4	7.92	0.43	3.1	20	0.93	0.13	8.62
	114-154	9.38	7.49	20.01	0.29	1.3	15.4	1.05	0.14	8.67
Bt,na C, na	154-200	9.4	7.52	16.26	0.16	2	12.2	1	0.11	8.62

**Table 2**  
Microbiological activity of virgin solonetz

	TNB (10 <sup>3</sup> g <sup>-1</sup> )	FNG (10 <sup>3</sup> g <sup>-1</sup> )	ACT (10 <sup>3</sup> g <sup>-1</sup> )	AMH (10 <sup>5</sup> g <sup>-1</sup> )	AZB (10 <sup>1</sup> g <sup>-1</sup> )	DHA (µg TPF/10g)
0-10 cm	74.2	31.24	14.19	69.03	98.94	661.9
10-20 cm	89.81	33.69	9.73	29.94	97.35	569.5
20-30 cm	68.95	36.31	15.15	22.04	79.76	502.8
average	77.65	33.75	13.02	40.34	92.02	578.1

**Table 3**  
**Influence of *Azotobacter* sp. and *Bacillus* sp. on microbiological activity in solonetz**

	TNB ( $10^6\text{g}^{-1}$ )	FNG ( $10^3\text{g}^{-1}$ )	ACT ( $10^4\text{g}^{-1}$ )	AMH ( $10^6\text{g}^{-1}$ )	AZB ( $10^5\text{g}^{-1}$ )	DHA ( $\mu\text{g TPF}/10\text{g}$ )
Control	7.8	33.8	1.3	4.0	0.0	578.1
A1	155.4	87.0	174.9	171.8	330.3	689.5
A2	223.0	58.1	164.1	201.3	329.0	690.8
A3	298.9	95.9	198.2	1153.5	351.3	715.5
A4	256.1	88.7	230.1	418.1	360.6	695.8
B1	131.4	74.3	165.9	104.1	330.6	655.9
B2	219.3	75.4	262.0	76.9	331.2	720.4

Note: A1-A3 isolates of *Azotobacter* sp., B1-B2 isolates of *Bacillus* sp.

*bacter* and *Bacillus* isolates were isolated, cultured in pure culture and reintroduced in soil in high number ( $10^{10}$  CFU/ml of *Azotobacter* sp. and  $10^{12}$  CFU/ml of *Bacillus* sp., respectively).

Introduction of indigineous bacteria isolates from virgin solonetz led to the increase of all investigated number of microorganisms as well as dehydrogenase activity (Table 3).

## Discussion

Increased salinity/sodicity (EC/Na) in the rhizosphere affects root exudation and biochemical transformations by microorganisms (Ondrasek and Rengel, 2012). High clay and increased Na content on soil at Bt horizon leads to colloid peptization and alkaline reaction of soil (Belić, 1999). These sodic soils are thus unfavorable or less favorable for microbial growth and activity. Similar observation were reported by Li et al. (2006), who noticed significant negative correlations between soil EC and microbial biomass C, suggesting that salinity had an adverse effect on microbial biomass and activity.

Brankov et al. (2006) investigated the microbiological activity of several soil types in Banat. According to the results of that research, soils with higher humus and nitrogen content as chernozem, solonetz and humogley were characterized by increased number of bacteria, azotobacter and dehydrogenase activity. Research showed that the most important limiting factor for plant production in solonetz is physiologically active depth of 22 cm (Škorić et al., 1985). Jarak et al. (2012) pointed out that number of microorganisms varied and depended on soil type and profile depth. They investigated the microbial abundancy in five soils such were sandy soil, solonetz, black soil, chernozem and fluvisol from different sites. The number and diversity of nitrogen fixing cyanobacteria were considerably higher in sodic and hydromorphic soils. The highest number of bacteria was counted in surface layer (0-5 cm). These soils were not used for agriculture production so the majority of easily decomposing soil organic matter was placed in the top soil resulting in increased bacteria number.

Katai et al. (2011) found lesser quantity of microscopic fungi in salt effected soil while larger number of aerobic cellulose decomposing and nitrifying bacteria was found in the cultivated soils. They also stated that the enzymes activities were generally higher in the natural soils than in the cultivated ones. The authors measured higher number of microorganisms and more intensive microbial activity in the soil from higher exposure indicating the importance of the micro relief in the occurrence and activity of living organisms of soil.

Generally, the abundance of fungi in comparison to bacteria is lower even after bacterial inoculation of soil. This is in accordance with the results of Sardinha et al. (2003). The authors found that the fungal biomarker ergosterol and the basal respiration rate continuously decreased from the non-saline sites to the saline-sodic sites. Fungi have been reported to be more sensitive to salinisation than bacteria in acidic soils, but also in alkaline soils (Badran, 1994; Pankhurst et al., 2001).

## Conclusions

The total number of bacteria, number of fungi, actinomycetes, free-living nitrogen fixing bacteria and aminoheterotrophs were increased in relation to the control. Dehydrogenase activity was also higher after bacterial inoculation. This research highlighted the positive effect of *Azotobacter* sp. and *Bacillus* sp. application. Further research needs to be carried out in order to increase the benefits of microbial inoculation in semi controlled and field conditions.

## Acknowledgements

This research is part of a project TD31027 financed by the Ministry of Education and Science, Republic of Serbia.

## References

- Badran, R. A. M., 1994. Cellulolytic activity of some cellulose-decomposing fungi in salinized soils. *Acta Mycology*, **29**: 221-245.

- Belić, M.**, 1999. Uticaj Meliorativnih Merana Adsorptivni Kompleks Solonjeca, Doctoral dissertation, *Faculty of Agriculture*, Novi Sad, Serbia (Sr).
- Belić, M., M. Dimitrijević, V. Hadžić, S. Petrović and Lj. Nešić**, 2003. The effect of amelioration on solonetz properties and yield variability of different wheat genotypes. *Agroznanjē, Nauka-Tehnologija-Praksa*, **4** (2): 89-99 (Sr).
- Brankov, M., M. Ubavić, P. Sekulić and J. Vasin**, 2006. The content of trace elements and heavy metals in agricultural and non-agricultural soils in Banat. *Zbornikradova, NaučniinstitutzaratarstvoiPovrtarstvo*, **42**: 169-177 (Sr).
- Jarak, M., N. Mrkovački, D. Bjelić, D. Jošić, T. Hajnal-Jafari and D. Stamenov**, 2012. Effects of plant growth promoting rhizobacteria on maize in greenhouse and field trial. *Afr. J. Microbiol. Res.*, **6**: 5683-5690.
- Katai, J., Á. OlahZsuposne and I. Vago**, 2011. Comparative evaluation of soil types with different properties. *Research Journal of Agricultural Science*, **43** (3): 87-94.
- Kruzhilin, I. P. and L. A. Kazakova**, 2011. Combined amelioration of soils of solonetz complexes in the Lower Volga region. *Russian Agricultural Sciences*, **37** (1): 55-57.
- Lenhard, G.**, 1956. Die dehydrogenase aktivitat das bodeusald mass fur die mikroorganizmentätigkeitim Boden. *Z. Pflanze Ernährung. Dung Bodenkude*, **73**: 1-11.
- Li, X., F. Li, B. Singh, Z. Cui and Z. Rengel**, 2006. Decomposition of maize straw in saline soil. *Biol. Fertil. Soils*, **42**: 366-370.
- Ondrasek, G. and Z. Rengel**, 2012. The role of organic matter in trace elements bioavailability and toxicity, In: A. Parvaiz and M. N. V. Prasad (Eds.) *Abiotic Stress Responses in Plants: Metabolism to Productivity*, Springer.
- Pankhurst, C. E., S. Yu, B. G. Hawke and B. D. Harch**, 2001. Capacity of fatty acid profiles and substrate utilization patterns to describe differences in soil microbial communities associated with increased salinity or alkalinity at three locations in South Australia. *Biology and Fertility of Soils*, **33**: 204-217.
- Sardinha, M., T. Muller, H. Schmeisky and R. G. Joergensen**, 2003. Microbial performance in a temperate floodplain soil along a salinity gradient. *Applied Soil Ecology*, **23**: 237-244.
- Sekulić, P., Lj. Nešić, V. Hadžić, M. Belić, J. Vasin, M. Ubavić, D. Bogdanović, M. Čuvarđić, D. Dozet, M. Pucarević, N. Milošević, M. Jarak, S. Đurić, J. Ralev and T. Škorić-Zeremski**, 2005. Soil of Serbia as a resource for sustainable development, plenarnireferat. In: *Zemljištekao Resurs Održivo Razvoja*, XI KongresDPZSCG, Budva, pp. 18-37 (Sr).
- Škorić, A., G. Filipovski and M. Ćirić**, 1985. Classification of soil in Yugoslavia. *Akademija Nauka i Umjetnosti Bosne i Hercegovine*, Sarajevo (Sr).
- Thalman, A.**, 1968. Zurmethodik des bestmmung des dehydrogenase aktivitatimbodenmittels TTC. *Landiw.Forch.*, **21**: 249-258.
- Trolldenier, G.**, 1996. Plate count technique. In: Franz Schinner (Ed.), *Methods in Soil Biology*, Springer-Verlag Berlin, Heidelberg, pp. 20-26.
- United Nations, FAO**, 2006. World Reference Base for Soil Resources 2006. *World Soil Resources Reports*, No. 103, Rome. <http://ftp.fao.org/agl/agll/docs/wsr103e.pdf> pp. 69-100