

## Radioprotective action of resurrection plant *Haberlea rhodopensis* Friv. (Gesneriaceae) and role of flavonoids and phenolic acids

Svetlana Georgieva<sup>1</sup>, Deyana Gencheva<sup>1</sup>, Borislav Popov<sup>2</sup>, Neli Grozeva<sup>3</sup>, Maria Zhelyazkova<sup>1</sup>

<sup>1</sup>Trakia University, Faculty of Agriculture, Department of Genetics, Animal breeding and Reproduction, Stara Zagora 6000, Bulgaria

<sup>2</sup>Trakia University, Faculty of Medicine, Department of Molecular Biology, Genetics and Immunology, Stara Zagora 6000, Bulgaria

<sup>3</sup>Trakia University, Faculty of Agriculture, Department of Biology and Aquaculture, Stara Zagora 6000, Bulgaria

\*Corresponding author: svetlana.georgieva8888@abv.bg

### Abstract

Georgieva, S., Gencheva, D., Popov, B., Grozeva, N. & Zhelyazkova, M. (2019). Radioprotective action of resurrection plant *Haberlea rhodopensis* Friv. (Gesneriaceae) and role of flavonoids and phenolic acids. *Bulg. J. Agric. Sci.*, 25 (Suppl. 3), 158–168

*Haberlea rhodopensis* Friv. (Gesneriaceae) is a Balkan endemic and resurrection glacial relict plant which is distributed in Bulgaria (Rhodope Mountains, Sredna Gora Mt. and Central Balkan). Currently, there is a huge interest in *Haberlea rhodopensis*, and several scientific studies aimed at the isolation and identification of its active components as well as the investigation of the pharmacological effects and possibilities of the use of *Haberlea rhodopensis* as a medical plant have been conducted. The main effects of *Haberlea rhodopensis* include radioprotective, anti-mutagenic, antioxidant and anti-ageing properties. This review emphasises the radioprotective potential of *Haberlea rhodopensis* and focuses on the biological properties of its ingredients like flavonoid aglycones and glycosides as well as phenolic acids in relation to their capacity to capture free radicals and reduce oxidative stress. More research on animals and humans are needed for clarification of the mechanisms of action and the eventual side effects of *Haberlea rhodopensis* and its compounds as radioprotective agents.

**Keywords:** Resurrection plants, *Haberlea rhodopensis*, radioprotection, flavonoids, phenolic acids

### Introduction

It is widely acknowledged that ionizing radiation causes damage to living organisms and induces a wide range of lesions in somatic and germ cells that can bring about mutations, cell inactivation, hereditary diseases and cancer. Irradiation causes an overabundance of reactive oxygen species (ROS) and free radicals that damage integral macromolecules in the living cells (Cox et al., 1995). The most important target for ionizing radiation is genomic DNA. DNA changes appear as a result of direct ionization leading to injury of both the nucleobases and sugar or indirectly

through the generation of ROS (Rehmana et al., 2016). If the repair of DNA is incorrect it may result in mutations, as double strand breaks, cross-links, base modifications, chromosome aberrations, etc. Apart from DNA, ROS attack lipids and proteins and cause oxidation, membrane damage, enzyme inactivation and other disorders (Ghosh et al., 2018). Consequently, since exposure to radiation accidentally or in radiotherapy can produce acute and late effects, it is very important to protect normal tissue by the use of nontoxic and efficient radioprotectors. Although synthetic compounds such as RW-2127 exhibited good protective properties, their toxicity in optimal doses promoted the search for new less

toxic and more effective alternatives (Jageta, 2007). This is why, over the decades a large number of plants and phytochemicals were considered as radioprotectors on account of their pharmacological properties and low toxicity (Cheki et al., 2016; Zbikowska et al., 2016; Ghali et al., 2018).

### **Resurrection Plants**

Resurrection plants are a small group of species belonging to different botanical families. What is typical of them is the high sustainability to extreme conditions like drought or desiccation. They have the ability to survive drought for months and years after which, in the presence of water, they can recover and continue to develop normally (Alpert & Oliver 2002). The mechanism of safekeeping of resurrection plants has been studied by many investigators (Blomstedt et al., 1998; Challabathula et al., 2016; Van Buren et al., 2018) and it is widely believed that defence is provided by a variety of processes including accumulation of different types of sugars, proteins and other substances which work together to substitute water and stabilise the subcellular environment. Via transcriptome analysis it was revealed that the drought induces new genetic programming, which directed resources from growth and development to cell protection (Gechev et al., 2013). Their observations suggested that both constitutive and inducible processes work concomitantly for the persistence during severe desiccation.

*Haberlea rhodopensis* (Friv.) (Fig.1) is an endemic resurrection plant, within a very small group of poikilohydrate angiosperms, member of the family Gesneriaceae. It is a Balkan Peninsula relic plant that has emerged before the glacial period. Gechev et al., (2014) reported that *Haberlea rhodopensis* under conditions of drying and humidification revealed common genetic pathway with other desiccation



**Fig. 1.** Resurrection plant *Haberlea rhodopensis*

tolerant species and has unique genes that may contribute to its desiccation sustainability. *Haberlea rhodopensis* is a unique plant that may serve not only as a model for the study of revitalization mechanisms after surviving strong water loss and as a source of genes involved in regeneration but also as a medicinal plant due to the beneficial properties of the ingredients of its extracts (Gechev et al., 2014).

### **Chemical composition of *Haberlea rodopensis***

The chemical composition of the family Gesneriaceae was studied by many authors (Lowry, 1972; Francisco, 1995; Jensen, 1996; Cai et al., 2005; Djilianov et al., 2009) and, as a result, flavonoids, caffeoyl phenylethanoid glycosides, tannins, anthocyanins, zeaxanthin, and ascorbate from various genders of the family have been established.

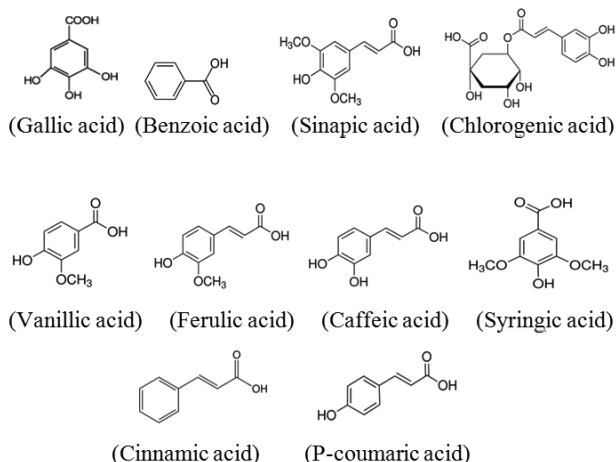
The availability of myconoside, paucifloside and 3 new flavone C-glycosides, hispidulin 8-C-(2''-O-syringoyl)-b-glucopyranoside, hispidulin 8-C-(6-O-acetyl-b-glucopyranoside), hispidulin 8-C-(6-O-acetyl-2-O-syringoyl-b-glucopyranoside), was reported in *Haberlea rhodopensis* by Ebrahimi et al., (2011). According to Berkov et al., (2011), metabolic profiling (gas chromatography in combination with mass spectrometry, GC-MS) of the polar and apolar fractions from methanolic extracts of *Haberlea rhodopensis*, showed availability of a large quantity of ingredients, including amino acids, fatty acids, several phenolic acids, sterols, glycerides, saccharides, flavonoids and polyphenols.

Microwave-assisted extraction of flavonoid antioxidants from *Haberlea rhodopensis* and evaluation for their total phenolic content also indicated the presence of flavonoid aglycones and glycosides (Fig. 3.) (Mihaylova et al., 2011). More than 100, mostly bioactive compounds have been found in *Haberlea rhodopensis* (Markovska et al., 1992; Djilianov et al., 2011; Moyankova et al., 2014). Recently the content of flavone C-glycosides and caffeoyl phenylethanoid glycosides in *Haberlea rhodopensis* was determined (Zheleva-Dimitrova et al., 2016) and it has been demonstrated that miconoside is the main compound in *Haberlea rhodopensis* extract ( $332.2 \pm 0.7$  mg/g dw), reaching to 88.8% from the estimated mixture in leaves, followed by paucifloside ( $24.8 \pm 2.1$  mg/g dw). Among the flavone 8-C-glycosides the most abundant was hispidulin 8-C-(6-O-acetyl-glucopyranoside ( $6.9 \pm 0.6$  mg/g dw) as the overall content of flavon C-glycosides was 17.1 mg/g dw.

### **Phenolic acids in *Haberlea rhodopensis* and their biological activities relevant to radiation protection**

According to many investigators, phenolic acids possess therapeutic properties that are due to their ability to capture free radicals and to decrease oxidative stress. They are widely used for the prophylaxis and treatment of cancer, diabetes,

inflammatory diseases and different oxidative stress-related diseases (Griffiths et al., 2016; Mut-Salud et al., 2016; Simoes et al., 2018). Mihaylova et al. (2011) reported that in the *Haberlea rhodopensis* alcoholic extract, the most abundant phenolic acids are ferulic acid (10.882 g/ml), caffeic acid (7.373 g/ml), p-coumaric acid (5.774 g/ml), etc. (Fig.2).



**Fig. 2. Chemical structure of phenolic acids in *Haberlea rhodopensis***

#### *Gallic acid (3,4,5-trihydroxybenzoic acid)*

The effects of gallic acid were studied under *in vitro* and *in vivo* circumstances and the results showed well defined radioprotective properties (Gandhi and Nair 2005). In their *in vitro* study, the authors tested the protective action of different concentrations of gallic acid in irradiated rat liver microsomes and plasmid pBR322 DNA. The results showed reduction of DNA injury and defence of microsomes against radiation-induced lipid peroxidation by gallic acid administrated during irradiation. At the same time, gallic acid was introduced before whole-body irradiation of mice, resulting in reduction of DNA damage in peripheral leucocytes and protection against lipid peroxidation *in vivo* (Gandhi & Nair, 2005).

Nair and Nair (2013) have also found some defensive action of gallic acid on lipid peroxidation in gamma-treated Swiss albino mice, along with raised levels of antioxidant enzymes, glutathione peroxidase and thiol glutathione. Additionally, they reported an anticlastogenic effect, activation of reparative processes in DNA and reduction of mortality and loss of weight in irradiated mice, as a result of gallic acid administration before irradiation.

Other conducted studies confirmed gallic acid as a proven antioxidant with neuroprotective (Mansouri et al., 2013), anticarcinogenic, antimutagenic and antiinflammatory action (Verma et al., 2013).

#### *Caffeic acid (3,4-dihydroxy)-cinnamic acid*

Numerous studies have indicated that caffeic acid is a strong antioxidant (Bors et al., 2004; Liu et al., 2019). Through different *in vitro* antioxidant assays Gülçin (2006) established that caffeic acid is a potent inhibitor of lipid peroxidation and manifested powerful radical scavenging capacity. A significant decrease of the density of chromosome aberrations, micronuclei and comet parameters was observed in human lymph cells administered with 66  $\mu$ M caffeic acid 30 min. before gamma irradiation (Devipriya et al., 2008).

Hakkim et al., (2014) have conducted an experiment with human skin cells to test the radioprotective action of 5 phenolic acids (caffeic acid, rosmarinic acid, trans-cinnamic acid, p-coumaric acid and hydroxyphenyllactic acid). The comparison of the results has shown that the introduction of caffeic acid, rosmarinic acid and trans-cinnamic acid before irradiation of keratinocytes improved the intracellular antioxidant balance and reduced DNA damage, as the most efficient was caffeic acid.

Sevgi et al., (2015) have explored the mutagenic action of UV and H<sub>2</sub>O<sub>2</sub> on plasmid BR322 DNA in the availability and unavailability of several phenolic acids, including caffeic acid. The results showed the defensive effect of caffeic acid against the DNA injury caused by the two mutagens. In addition, caffeic acid was found to be effective against inflammation, mutagenesis, bacterial infections and carcinogenesis, due to its radical scavenging capacity (Genaro-Mattos et al., 2015).

#### *Ferulic acid (4-hydroxy-3-methoxy cinnamic acid)*

Ferulic acid is known as a strong antioxidant. Those effects are due to the donation of electrons from hydroxyl and phenoxy groups of ferulic acid that scavenge the free radicals (Srinivasan et al., 2007). For a more detailed explanation of the effects of ferulic acid, Prasad et al., (2006) used 3 different concentrations of ferulic acid and 3 different doses for gamma irradiation of lymphocyte cultures. The results from this experiment showed that administration of ferulic acid prior to exposure of human lymph cells, significantly reduced DNA damage as indicated by the reduced quantity of micronuclei, dicentric aberration and lipid peroxidation and also by the improved antioxidant condition of the cells.

In another experiment, mice were treated with various quantities of ferulic acid (50, 75 and 100 mg/kg body weight) one hour prior exposure to 4.0Gy whole body gamma irradiation. The introduction of ferulic acid induced a concentration-dependent reduction in the DNA injuries in peripheral blood leukocytes and bone marrow cells of mice (Maurya et al., 2005).

Similar evidence for radioprotective action of ferulic acid is presented by Shanthakumar et al., (2012). Their study *in vivo* with irradiated Swiss albino mice indicated that pre-treatment with ferulic acid resulted in decrease of inflicted lipid peroxidation and DNA defects, and also improved antioxidant and histopathological amendments in animals. They also reported an antigenotoxic result of ferulic acid *in vitro* expressed by a significant decrease of the comet parameters (% tail DNA, tail length, tail moment and Olive tail moment, comet assay).

#### *P-coumaric acid (4-hydroxycinnamic acid)*

Lodovici et al., (2001) investigated the effect of several native phenolic acids on DNA oxidation *in vitro* and found that p-coumaric acid diminished DNA injury caused by Fe and cumene hydroperoxide.

In another *in vivo* experiment, rats were treated for 2 weeks with p-coumaric acid in the diet (25 or 50 mg/kg) and results revealed that p-coumaric acid (50 mg/kg) decreased effectively the basic level of oxidative DNA injury in rat colonic mucosa (Guglielmi et al., 2003). Because of its powerful anti-oxidant and anti-apoptotic potential p-coumaric acid manifested neuroprotective action in a rat model of embolic cerebral ischemia (Güven et al., 2015). P-coumaric acid effectively scavenges free radicals, diminishes lipid peroxidation, and also possesses antibacterial, antimutagenic and immunoregulatory activities (Andreicutu et al., 2019; Shen et al., 2019).

#### *Chlorogenic acid - 3-(3,4-dihydroxycinnamoyl)quinic acid*

Cinkilic et al. (2013) investigated *in vitro* by comet assay the safeguarding effect of different quantities of chlorogenic acid administered in human lymphocytes in the presence and lack of X radiation. They observed a lack of genotoxicity in lymphocytes treated with chlorogenic acid only and effective protection against X-ray induced genomic instability. The most effective dose of chlorogenic acid, diminishing by 48.15% the DNA damage was 4 µg/ml. Other authors in the experiments with gamma-irradiated mouse bone marrow cells and human lymphocytes reported that chlorogenic acid acts as a strong antioxidant, free radical neutralizer and anticarcinogen (Hosseinimehr et al., 2007; Cheki et al., 2016). Likewise, it was found that chlorogenic acid inhibited MAM acetate-caused carcinogenesis in colon and liver of hamsters (Mori et al., 1986).

#### *Sinapic acid (3,5-dimethoxy-4-hydroxycinnamic acid)*

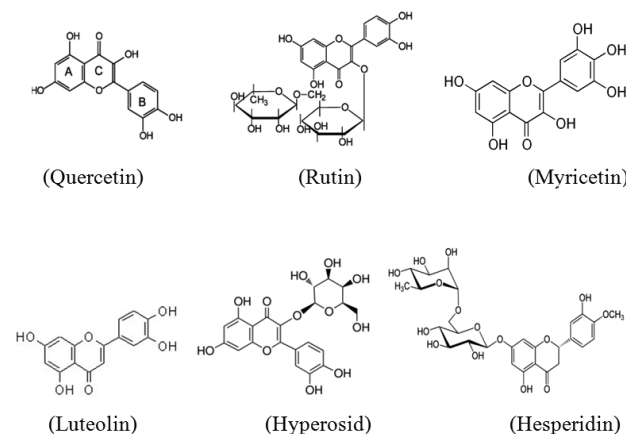
Sinapic acid is revealed to have strong antioxidant capacity and free radical scavenging potential to catch superoxide anion ( $O_2^-$ ), hydroxyl radicals ( $\cdot OH$ ) and other free

radicals (Chen, 2016). According to Thiyam et al., (2006) sinapic acid has the potential to inhibit the products of lipid peroxidation. Hameed et al., (2016) analysed the usefulness for humans of the sinapic acid and its derivatives, particularly 4-vinylsyringol, and established safeguarding effect of sinapic acid against inflammation, mutagenesis and cancer.

#### *Cinnamic acid (3-phenylacrylic acid)*

The radioprotective effectiveness of cinnamic acid has been tested in X-ray irradiated human blood lymph cells *in vitro*. Initially, in this experiment it was found out that cinnamic acid did not exhibit genotoxic activity on non-irradiated lymphocytes and then it was revealed that cinnamic acid administered to irradiated lymphocytes cultures decreased DNA injuries caused by X-rays, as indicated by a reduced quantity of micronuclei (16-55%) and DNA breaks (17-50%) in contrast with irradiated lymphocytes (Cinkilic et al., 2014). Sova (2012) also reported that cinnamic acid has low toxicity and a broad spectrum of biological activities, due to its strong ability to capture free radicals.

As we have already mentioned, the extraction of flavonoid antioxidants from *Haberlea rhodopensis* and evaluation of their total phenolic content was shown the presence of significant quantities of flavonoid aglycones and glycosides (Fig. 3) (Mihaylova et al., 2011).



**Fig. 3. Flavonoid aglycones and glycosides in *Haberlea rhodopensis***

According to Mihaylova et al., (2011) among the flavonoid aglycones and glycosides in *Haberlea rhodopensis* the most abundant is luteolin (55.118 g/ml) followed by hesperidin (10.122 g/ml), myricetin (11.7 g/ml), rutin, quercetin, etc.

***Flavonoid aglycones and glycosides in *Haberlea rhodopensis* and their biological activities relevant to radiation protection***

*Luteolin (3',4',5,7-tetrahydroxyflavone)*

Early experimental research of Shimoi et al. (1996) have shown strong anticlastogenic and antioxidative effect of luteolin. In this article luteolin was revealed as the most effective anticlastogen (micronucleus test) among the 12 tested flavonoids from rooibos tea (*Aspalathus linearis*). In addition, gastric administration of luteolin before high dose gamma irradiation led to an increase of endogenous antioxidants and a decrease of lipid peroxidation in mouse bone marrow and spleen. Anticlastogenic activity of luteolin against X-rays was also established in mouse bone marrow cells by Benavente-García et al., (2004).

The flavone luteolin exhibited a high protective effect against the t-BHP-induced DNA strand breaks with a reduction in DNA damage, evaluated by the Comet assay (Silva, et al., 2008). Materska et al. (2015) reported that luteolin manifested radioprotective activity against X-rays. They also determined the high coefficient of the correlation between the X-rays radioprotective effect of phenolic glycosides and their scavenging activities against O<sub>2</sub><sup>-</sup> generated in the NADH/PMS system.

Lien et al. (1999) reported that luteolin acts against ROS through its own oxidation, and restricts ROS-generating oxidases (Nagao et al., 1999).

Many other investigators have established that luteolin has a multiplicity of medical properties, including antioxidant, antimicrobial, anti-inflammatory, anti-cancer activity and possibilities to enhance endogenous antioxidants (Leung et al., 2006; Lin et al., 2008; Moga et al., 2016).

*Hesperidin (4'-methoxy-3',5,7-trihydroxyflavanone)*

Studies have shown the antioxidant potential of hesperidin and its safeguarding action against H<sub>2</sub>O<sub>2</sub>-induced oxidative damage on pBR322 DNA and shielding effect to human lymph cells and RBC cellular membranes against radiation-induced damage (Kalpana et al., 2009 a,b).

The protective effect of IP administration of hesperidin in different doses prior to gamma irradiation (2 Gy) of mice was proven by the dose-dependent reduction of radiation-induced DNA damage in PCEs of mice (Hosseinimehr & Nemati 2006). Hesperidin was also found to reduce oxidative stress and decrease hepatocellular damage in Sprague-Dawley rats (Pradeep et al., 2008).

The positive effects of hesperidin were evaluated in experimental animals by different methods: 30 days of survival study, biochemical assay, comet assay, DNA fragmentation

assay and histopathological alterations in the mouse liver (Kalpana et al., 2011). The authors established an antioxidant effect, defence against DNA damage and protection against liver damage in irradiated mice, pre-treated with hesperidin.

In general, hesperidin has been proven to have a broad spectrum of pharmacological effects, which also include anti-inflammatory, anti-allergic, hypolipidemic, vasoprotective and anti-carcinogenic action (Emin et al., 1994; Haddadi et al., 2017).

*Myricetin (3,3',4',5,5',7-hexahydroxyflavone)*

Different investigations have indicated that myricetin has an antioxidant, anti-inflammatory and efficient anticancer effect (Ross & Kasum, 2002; Sun et al., 2012; Semwal et al., 2016; Arruda et al., 2018; Jarijala et al., 2018). Aherne & O'Brien (1999) in their study examined the effect of the 3 flavonoids – myricetin, quercetin, and rutin, on DNA *in vitro*. They used Caco-2 and Hep G2 cell lines exposed to H<sub>2</sub>O<sub>2</sub>. After pre-incubation with myricetin and the other 2 flavonoids followed by H<sub>2</sub>O<sub>2</sub> exposure of cell lines, they found a significant diminution of DNA injury in both cell lines.

Abalea et al., (1999) also investigated the protective effect of different concentrations of myricetin against iron-induced DNA damage in cultures of rat hepatocytes. They reported that simultaneous administration of myricetin and iron protects against DNA base oxidation and activates the expression of DNA repair enzymes in a dose-dependent manner.

*Quercetin (3,3',4',5,7-pentahydroxyflavone) and rutin (3,3',4',5,7-pentahydroxyflavone-3-rutinoside)*

Lots of experimental outcomes revealed that quercetin and its derivative rutin are excellent *in vitro* antioxidants. The antioxidant and radiodefensive activity of both flavonoids was investigated in mice treated with gamma radiation by Patil et al., (2013). The results revealed that the application of rutin and quercetin caused elevation of endogenous antioxidants, reduction of lipid peroxidation and inhibition of various free radicals in a dose-dependent way. They also determined radical scavenging activity *in vitro* by different assays (2,2-diphenyl-1-picrylhydrazyl (DPPH), O<sub>2</sub>, 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid) (ABTS)<sup>+</sup>, and OH) and concluded that the radioprotective action of rutin and quercetin is probably due to their antioxidant features and high ability to capture free radicals.

In agreement with the previous authors Londhe et al., (2009) reported the defensive effect of rutin and quercetin expressed through diminished lipid peroxidation and protein oxidation in rat liver mitochondria. They also established reduction of radiation-induced DNA damage and more specifi-

cally diminishing in single-strand breaks in plasmid DNA. According to those authors the described effects can be explained by the strong hydroxyl radical scavenging activity of the phytochemicals.

The antigenotoxicity of rutin and quercetin was demonstrated by Patil et al., (2014). In their investigation the introduction of rutin and quercetin before exposure led to a significant diminution in the chromosome aberrations in mice bone marrow cells, micronuclei in PCE and olive tail movement (OTM) in blood leukocytes and those effects are an indicator for anticlastogenicity and antigenotoxicity of rutin and quercetin.

The antigenotoxic effect of quercetin and rutin was also proven on hep G2 cells exposed to H<sub>2</sub>O<sub>2</sub> by Barcelos et al., (2011). Some protection against H<sub>2</sub>O<sub>2</sub> -induced oxidative stress was found by the administration of both phytochemicals at low non-toxic doses, although at higher concentrations an increase in intracellular ROS was observed.

According to Silva et al., (2008) quercetin protected DNA against oxidative damage of PC12 cells, as the repair of DNA reached 24.7%. In an *in vivo* experiment, the defence by quercetin against radiation-induced DNA injury and apoptosis in kidney and other tissues of rats was proven by Özyurt et al. (2014). Sancar et al., (2004) reported a protective effect of quercetin against DNA strand breaks, enhancement of the repair of MMS-induced damage and excision of damaged nucleotides. Quercetin also showed high radioprotective action (50% vs control) on human lymph cells in response to X ray-induced oxidative damage (Materska et al., 2015) and reduction of percentage mortality and cytogenetic damage in gamma irradiated Swiss mice (Patil et al., 2014).

The anti-cancer activity of quercetin has been examined *in vitro* and *in vivo* by Gibellini et al., (2010) and Hashemzai et al., (2017). Quercetin at the proper concentrations was found to induce the apoptosis of some tested cancer cell lines and to cause a significant reduction in tumour volume in mice bearing tumours. In addition, quercetin depleted intracellular glutathione and increased intracellular ROS in cancer cells to a level that can cause cell death.

According to Kondareva-Burdina et al. (2013) along with the above mentioned compounds of *Haberlea rhodopensis* with useful properties, the caffeoyl phenyl-ethanoid glycoside myconoside and flavone-C-glycosides hispidulin 8-C-(2-O-syringoyl-gluco-pyranoside), hispidulin 8-C-(6-O-acetyl-2-O-syringoyl-gluco-pyranoside), and hispidulin 8-C-(6-O-acetyl-gluco-pyranoside) isolated from *Haberlea rhodopensis* have shown a good relationship between the cyto-protection of rat hepatocytes, free radical scavenging and antioxidant effect. The authors also reported that myconoside possesses the strongest DPPH radical scavenging and antioxidant potential in the linoleic acid system.

#### **Radioprotective effects of *Haberlea rhodopensis* extract**

A number of medicinal plants and their ingredients have been studied and their radioprotective effect was established *in vivo* and *in vitro* (Taheri et al., 2016; Chandrasekara & Shahidi, 2018; Shaikh & Shah, 2018). The results showed that plant extracts that exhibit radioprotective effects contain a variety of compounds with antioxidants, free radical scavenging, anticlastogenic, antigenotoxic, antimicrobial, and immunomodulatory properties.

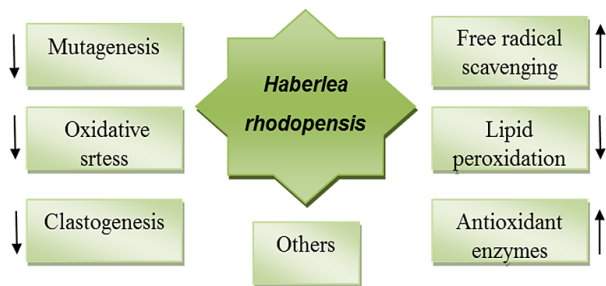
In our studies, the *Haberlea rhodopensis* leaf extract has been tested as an antioxidant and radioprotector in experimental animals - New Zealand rabbits. The radioprotective property of *Haberlea rhodopensis in vitro* was first reported by Popov et al. (2010). The effect of *Haberlea rhodopensis* was investigated in the blood lymphocytes of rabbits after exposure to gamma irradiation and the results showed that *Haberlea rhodopensis* pre-treatment of lymphocyte cultures effectively diminished the quantity of aberrant cells and chromosome aberrations. The comparison of the anticlastogenic effect of *Haberlea rhodopensis* with vitamin C indicated that both the reduction of aberrant cells, the density of chromosome fragments and dicentrics are highly expressed after *Haberlea rhodopensis* treatment.

In other *in vivo/in vitro* studies, the genetic deteriorations in peripheral blood lymphocytes exposed to gamma rays (1.0-3.0 Gy) indicated that radiation-induced increase of chromosome aberrations is dose-dependent. Pre-treatment with different concentrations of *Haberlea rhodopensis* extract strongly diminished the quantity of aberrant cells and chromosomal aberrations, including dicentrics and rings, in a dose-dependent manner (Popov et al., 2011; 2012; 2013).

The alkaline single cell gel electrophoresis analysis of lymph cells from the whole body irradiated rabbits, pre-treated with *Haberlea rhodopensis* indicated a reduction in induced cellular DNA damage, like the formation of alkali-labile sites and single and double breaks (Georgieva et al., 2012).

In addition, a biochemical and cytogenetic assay has been carried out to detect the modifying effect of *Haberlea rhodopensis* against radiation-induced oxidative stress. It was observed that *Haberlea rhodopensis* pre-treatment significantly increased the activity of the antioxidant enzymes SOD and CAT (compared between the control and irradiated groups). At the same time the anti-lipid peroxidative effect of *Haberlea rhodopensis* was registered through the reduction of MDA level in blood plasma (Georgieva et al., 2013). Furthermore, cytokinesis blocked micronucleus assay (CBMN) in peripheral lymphocytes revealed a significant reduction of the quantity of MN events in lymphocytes protected with *Haberlea rhodopensis* (Georgieva, 2014).

These observations and the information about the compounds in *Haberlea rhoropensis* and their biological activities relevant to radiation protection provide information on the mechanism of the action of the extract. Fig.4



**Fig. 4. Mechanism of action of *Haberlea rhoropensis* extract**

Many authors have reported a linear correlation between the content of all phenolic compounds and their antioxidant capacity (Katalinic et al., 2006; Chen et al., 2016; Srirachoen et al., 2017; Verma et al., 2018).

The results obtained for *Haberlea rhoropensis* evaluated by reducing power assay activity showed a good positive correlation  $R^2$  0.8286, which suggested that the major antioxidant activity was due to phenolic compounds, which determined its pharmacological properties (Michaylova et al., 2011). In addition to that, to evaluate the usefulness of *Haberlea rhoropensis* for application in phytotherapy Berkov et al., (2011) investigated the cellular viability and cell survival of five cell lines and a lack of any toxicity in tested lines has been found.

## Conclusion

The possibilities of using the extracts of medicinal plants, including *Haberlea rhoropensis*, depend on their phytochemical spectrum. Polyphenols such as phenolic acids, flavonoid aglycones and glycosides manifest their action through complex mechanisms. They may capture free radicals, improve the antioxidant balance, regulate the genes responsible for DNA repair and influence the expression of mRNAs of antioxidant enzymes, thus restoring the disequilibrium in cells exposed to ionizing radiation. The presence of an intense antioxidant, free radical scavenging and radioprotective potential as well as the lack of cytotoxic activity of *Haberlea rhoropensis* indicates that it may be used in phytotherapy and radiotherapy, but only after further research has been done and when the mechanisms of its action are basically studied.

## References

- Abalea, V., Cillard, J., Dubos, M. P., Sergent, O., Cillard, P. & Morel, I. (1999). Repair of iron-induced DNA oxidation by the flavonoid myricetin in primary rat hepatocyte cultures. *Free Radical Biology and Medicine*, 26, 1457-1466.
- Aherne, S.A. & O'Brien, N. M. (1999). Protection by the flavonoids myricetin, quercetin, and rutin against hydrogen peroxide-induced DNA damage in Caco-2 and Hep G2 cells. *Nutrition and Cancer*, 34, 160-166.
- Alpert, P. & Oliver, M. (2002). Drying without dying, in: Desiccation and Survival in Plants: Drying without Dying. Black M., Pritchard H. (eds), New York, NY: CABI Publishing, 1-43.
- Andreicuț, A. D., Pârnu, A.E., Moț, A. C., Parvu, M., Fischer-Fodor, E., Feldrihan, V., Cătoi, A. F., Cecan, M. & Irimie, A. (2019). Anti-Inflammatory and antioxidant effects of Mahonia aquifolium leaves and bark extracts. *Farmacia*, 66: 49-58.
- Arruda, H. S., Pereira, G. A. & Pastore, G. M. (2018). Brazilian Cerrado fruit araticum (*Annona crassiflora* Mart.) as a potential source of natural antioxidant compounds. *International Food Research Journal*, 25(5), 2005-2012.
- Barcelos, G.R.M., Grotto, D., Angeli, J.P.F., Serpeloni, J.M., Rocha, B.A., Bastos, J.K & Barbosa, F.J. (2011). Evaluation of Antigenotoxic effects of plant flavonoids quercetin and rutin on HepG2 cells. *Phytotherapy Research*, 25, 1381-1388.
- Benavente-García, O., Castillo, J., Lorente, J. & Alcaraz, M. (2004). Radioprotective Effects In Vivo of Phenolics Extracted from *Olea europaea* L. Leaves Against X-Ray-Induced Chromosomal Damage: Comparative Study Versus Several Flavonoids and Sulfur-Containing Compounds. *Journal of Medicinal Food*, 5, 125-135.
- Berkov, S. H., Nikolova, M. T., Hristozova, N., Momekov, G., Ionkova, I. I. & Djilianov, D. (2011). GC-MS profiling of bioactive extracts from *Haberlea rhoropensis*: an endemic resurrection plant. *Journal of Serbian Chemical Society*, 76, 211-220.
- Blomstedt, C. K., Gianello, R. D., Gaff, D. F., Hamill, J. D. & Neale, A. D. (1998). Differential gene expression in desiccation-tolerant and desiccation sensitive tissue of the resurrection grass *Sporobolus stapfianus*. *Australian Journal of Plant Physiology*, 25, 937 - 946.
- Bors, W., Michel, C., Stettmaier, K., Lu, Y. & Foo, L. Y. (2004). Antioxidant mechanisms of polyphenolic caffeic acid oligomers, constituents of *Salvia officinalis*. *Biological Research*, 37, 301-311.
- Cai, X. H., Luo, X. D., Zhou, J. & Hao, X. J. (2005). Quinones from *Chirita eburnea*. *Journal of Natural Products*, 68, 797-799.
- Challabathula, D., Puthur, J. T. & Bartels, D. (2016). Surviving metabolic arrest: photosynthesis during desiccation and rehydration in resurrection plants. *Annals of the New York Academy of Sciences*, 1365(1), 89-99.
- Chandrasekara, A. & Shahidi, F. (2018). Herbal beverages: Bioactive compounds and their role in disease risk reduction - A review. *Journal of Traditional and Complementary Medicine*, 8, 451-458.

- Cheki, M., Mihanooost, E., Shirazi, A & Mahmoudzadeh, A.** (2016). Prophylactic role of some plants and phytochemicals against radio-genotoxicity in human lymphocytes. *Journal of Cancer Research and Therapeutics*, 12, 1234-42.
- Chen, C.** (2016). Sinapic acid and its derivatives as medicine in oxidative stress-induced diseases and aging. *Oxidative Medicine and Cellular Longevity*, 3571614:1-10.
- Chen, G.L., Chen, S. G., Xie, Y. Q., Chen, F., Zhao, Y. Y., Luo, C.X. & Gao, Y. Q.** (2015). Total phenolic, flavonoid and antioxidant activity of 23 edible flowers subjected to in vitro digestion. *Journal of Functional Foods*, 17, 243-259.
- Cinkilic, N., Cetintas, S. K., Zorlu, T., Vatan, O., Yilmaz, D., Cavas, T., Tunc, S., Ozkan, L. & Bilaloglu, R.** (2013). Radioprotection by two phenolic compounds: Chlorogenic and quinic acid, on X-ray induced DNA damage in human blood lymphocytes in vitro. *Food and Chemical Toxicology*, 53, 359-363.
- Cinkilic, N., Tüzün, E., Çetintaş, S.K., Vatan, Ö., Yılmaz, D., Çavaş, T., Tunc, S., Özkan, L. & Bilaloğlu, R.** (2014). Radioprotective effect of cinnamic acid, a phenolic phytochemical, on genomic instability induced by X-rays in human blood lymphocytes in vitro. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 770, 72-79.
- Cox, J. D., Stetz, J. & Pajak, T. F.** (1995). Toxicity criteria of the Radiation Therapy Oncology Group (RTOG) and the European Organization for Research and Treatment of Cancer (EORTC). *International Journal of Radiation Oncology, Biology, Physics*, 31, 1341-1346.
- Devipriya, N., Sudheer, A. R. & Menon, V. P.** (2008). Caffeic acid protects human peripheral blood lymphocytes against gamma radiation-induced cellular damage. *Journal of Biochemical and Molecular Toxicology*, 22, 175-186.
- Djiljanov, D., Ivanov, S., Georgieva, T., Moyankova, D., Berkov, S., Petrova, G., Mladenov, P., Christov, N., Hristozova, N., Peshev, D., Tchorbadijeva, M., Alexieva, V., Tosheva, A., Nikolova, M., Ionkova, I. & Van den Ende, W.** (2009). A holistic approach to resurrection plants *Haberlea rhodopensis* - a case study. *Biotechnology and Biotechnological Equipment*, 23, 1414-1416.
- Djiljanov, D., Ivanov, S., Moyankova, D., Miteva, L., Kirova, E., Alexieva, V., Joudi, M., Peshev, D. & Van den Ende, W.** (2011). Sugar ratios, glutathione redox status and phenols in the resurrection species *Haberlea rhodopensis* and the closely related non-resurrection species. *Chirita eberhardtii* plant biology, 13, 767-776.
- Ebrahimi, N. S., Gafner, F., Acquac, G. D., Schweikert, K. & Hamburger M.** (2011). Flavone 8-C-glycosides from *Haberlea rhodopensis* Friv. (Gesneriaceae). *Helvetica Chimica Acta*, 94, 38 - 45.
- Emin, J.A., Oliveira, A.B. & Lapa, A. J.** (1994). Pharmacological evaluation of the antiinflammatory activity of a citrus bioflavonoid, hesperidin and the isoflavonoids dauricin and clausenquinone in rats and mice *The Journal of Pharmacy and Pharmacology*, 46, 118-122.
- Francisco, A.** (1995). The flavonoids—Advances in research since 1986. J.B. Harborne (Ed.) Chapman & Hall, London, UK 1994, pp.676. *Phytochemical Analysis* 6(1): 55-55.
- Gandhi, N. M. & Nair, C. K.** (2005). Protection of DNA and membrane from gamma radiation induced damage by gallic acid. *Molecular and Cellular Biochemistry*, 278, 111-117.
- Gechev, T. S., Benina, M., Obata, T., Tohge, T., Sujeeth, N., Minkov, I., Hille, J., Temanni, M. R., Marriott, A. S., Bergström, E., Thomas-Oates, J., Antonio, C., Mueller-Roeber, B., Schippers, J. H., Fernie, A. R. & Toneva, V.** (2013). Molecular mechanisms of desiccation tolerance in the resurrection glacial relic *Haberlea rhodopensis*. *Cellular and Molecular Life Sciences*, 70, 689-709.
- Gechev, T. S., Hille, J., Woerdenbag, H.J., Benina, M., Mehterov, N., Toneva, V., Fernie, A. R. & Mueller-Roeber, B.** (2014). Natural products from resurrection plants: Potential for medical applications. *Biotechnology Advances*, 32, 1091-1101.
- Genaro-Mattos, T. C., Maurício, Â. Q., Rettori, D., Alonso, A. & Hermes-Lima, M.** (2015). Antioxidant Activity of Caffeic Acid against Iron-Induced Free Radical Generation -A Chemical Approach. *PLOS ONE*, 10, 1-12.
- Georgieva, S.** (2014). Evaluation of Radiomodulatory Effect of *Haberlea Rhodopensis* (Friv.) Extract Using Micronucleus Assay in Lymphocytes of Whole Body Irradiated Rabbits. *Indian Journal of Applied Research*, 4, 6-8.
- Georgieva, S., Popov, B. & Bonev, G.** (2013). Radioprotective effect of *Haberlea rhodopensis* (Friv.) leaf extract on  $\gamma$ -radiation-induced DNA damage, lipid peroxidation and antioxidant levels in rabbit blood. *Indian Journal of Experimental Biology*, 51, 29-36.
- Georgieva, S., Popov, B., Milochev, G. & Bonev, G.** (2012). Cellular DNA damage and lipid peroxidation in rabbits after whole body gamma irradiation and treatment with *Haberlea rhodopensis* extract in rabbits. *Revue de Medecine Veterinaire*, 163, 572-576.
- Ghali, E. N. H. K., Maurya, D. K. & Meriga, B.** (2018). Radioprotective Properties of *Pterocarpus santalinus* Chloroform Extract in Murine Splenic Lymphocytes and Possible Mechanism. *Cancer Biotherapy and Radiopharmaceuticals*, 33, 10, <https://doi.org/10.1089/cbr.2018.2532>
- Ghosh, N., Das, A., Chaffee, S., Roy, S., & Sen, S.K.** (2018). Chapter 4 - Reactive Oxygen Species, Oxidative Damage and Cell Death. Immunity and Inflammation in Health and Disease. Emerging Roles of Nutraceuticals and Functional Foods in Immune Support 2018: 45-55.
- Gibellini, L., Pinti, M., Nasi, M., De Biasi, S., Roat, E., Bertonecelli, L. & Cossarizza, A.** (2010). Interfering with ROS Metabolism in Cancer Cells: *The Potential Role of Quercetin* *Cancers*, 2, 1288-1311.
- Griffiths, K., Aggarwal, B. B., Singh, R. B., Buttar, H. S., Wilson, D. & De Meester, F.** (2016). Food Antioxidants and Their Anti-Inflammatory Properties: A Potential Role in Cardiovascular Diseases and Cancer Prevention. *Diseases*, 4, 28.1-15,
- Guglielmi, F., Luceri, C., Giovannelli, L., Dolara, P. & Lodovici, M.** (2003). Effect of 4-coumaric and 3,4-dihydroxybenzoic acid on oxidative DNA damage in rat colonic mucosa. *British Journal of Nutrition*, 89, 581-587.
- Güven, M., Aras, A.B., Akman, T., Sen, H.M., Ozkan, A., Saliş, O., Sehitoglu, I., Kalkan, Y.C., Deniz, M. & Cosar, M.** (2015). Neuroprotective effect of p-coumaric acid in rat model of embolic cerebral ischemia. *Iranian journal of basic medical*



- sciences*, 18, 356-363.
- Gülçin, I.** (2006). Antioxidant activity of caffeic acid (3,4-dihydroxycinnamic acid). *Toxicology*, 217, 213-220.
- Haddadi, G. H; Rezaeyan, A., Mosleh-Shirazi, M. A., Hosseinzadeh, M., Fardid, R., Masoud Najafi, M & Salajegheh,** (2017). A. Hesperidin as Radioprotector against Radiation-induced Lung Damage in Rat: A Histopathological Study. *Journal of Medical Physics*, 42, 25–32.
- Hakim, F. L., Miura, M., Matsuda, N., Alharassi, A.S., Guillemin, G., Yamauchi, M. & Song, H.** (2014). An in vitro evidence for caffeic acid, rosmarinic acid and trans cinnamic acid as a skin protectant against  $\gamma$ -radiation. *International Journal of Low Radiation*, 9, 305 - 316.
- Hameed, H., Aydin, S. & Başaran, N.** (2016). Sinapic Acid: Is It Safe for Humans? *FABAD Journal of Pharmaceutical Sciences*, 41, 39-49.
- Hashemzaei, M., Far, A. D., Yari, A., Heravi, R. E., Tabrizian, K., Taghdisi, S. M., Sadegh, S.E., Tsarouhas, K., Kouretas, D., Tzanakakis, G., Nikitovic, D., Anisimov, N. Y., Spandidos, D. A., Tsatsakis, A. & Rezaee, R.** (2017). Anticancer and apoptosis-inducing effects of quercetin *in vitro* and *in vivo*. *Oncology Reports*, 38, 819-828.
- Hosseinimehr, S. J. & Nemat, A.** (2006). Radioprotective effects of hesperidin against gamma irradiation in mouse bone marrow cells. *The British Journal of Radiology*, 79, 415-418.
- Hosseinimehr, S. J., Azadbakht, M., Mousavi, S.M., Mahmoudzadeh, A. & Akhlaghpour, S.** (2007). Radioprotective effects of hawthorn fruit extract against gamma irradiation in mouse bone marrow cells. *Journal of Radiation Research*, 48, 63-68.
- Jagetia, G. C.** (2007). Radioprotective Potential of Plants and Herbs against the Effects of Ionizing Radiation. *Journal of clinical biochemistry and nutrition*, 40(2), 74-81.
- Jariala, R., Thakura, S., Sakinaha, M., Zularisama, A.W.Z., Sharadb, A., Kanware, S.S. & Singha L.** (2018). Potent anticancer, antioxidant and antibacterial activities of isolated flavonoids from *Asplenium nidus*. *Journal of King Saud University – Science*, 30, 185-192.
- Jensen, S. R.** (1996). Phenylethanoid glycosides in *Sanango racemosa* and in the family *Gesneriaceae*. *Phytochemistry*, 43, 777-787.
- Kalpana, K.B., Devipriya, N., Srinivasan, A., Vishwanathan, P., Thayalan, M. & Menon, V.P.** (2011). Evaluating the radioprotective effect of hesperidin in the liver of Swiss albino mice. *European Journal of Pharmacology*, 658, 206-212.
- Kalpana, K. B., Devipriya, N., Srinivasan, M. & Menon, V.P.** (2009). Investigation of the radioprotective efficacy of hesperidin against gamma-radiation induced cellular damage in cultured human peripheral blood lymphocytes. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 676, 54-61.
- Kalpana, K.B., Srinivasan, M. & Menon, V.P.** (2009). Evaluation of antioxidant activity of hesperidin and its protective effect on H<sub>2</sub>O<sub>2</sub> induced oxidative damage on pBR322 DNA and RBC cellular membrane. *Molecular and Cellular Biochemistry*, 323, 21-29.
- Katalinic, V., Milos, M., Kulisic, T. & Jukic, M.** (2006). Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols. *Food Chemistry*, 94, 550-557.
- Kondeva-Burdina, M., Zheleva-Dimitrova, D., Nedialkov, P., Girreiser, U. & Mitcheva, M.** (2013). Cytoprotective and antioxidant effects of phenolic compounds from *Haberlea rhodopensis* Friv. (Gesneriaceae). *Pharmacognosy Magazine*, 9, 294-301.
- Leung, H. W., Kuo, C. L., Yang, W. H., Lin, C. H. & Lee, H. Z.** (2006). Antioxidant enzymes activity involvement in luteolin-induced human lung squamous carcinoma CH27 cell apoptosis. *European Journal of Pharmacology*, 534, 12-18.
- Lien, E. J., Ren, S., Bui, H.H. & Wang, R.** (1999). Quantitative structure-activity relationship analysis of phenolic antioxidants. *Free Radical Biology and Medicine*, 26, 285-294.
- Lin, Y., Shi, R., Wang, X. & Shen, H. M.** (2008). Luteolin, a flavonoid with potentials for cancer prevention and therapy. *Current Cancer Drug Targets*, 8, 634-646.
- Liu, Q., Tang, G.Y., Zhao, C.N., Gan, R.Y. & Li, H. B.** (2019). Antioxidant Activities, Phenolic Profiles, and Organic Acid Contents of Fruit Vinegars. *Antioxidants*, 8, 78-90.
- Lodovici, M., Guglielmi, F., Meoni, M. & Dolara, P.** (2001). Effect of natural phenolic acids on DNA oxidation *in vitro*. *Food and Chemical Toxicology*, 39, 1205-1210.
- Londhe, J. S., Devasagayam, T. P. A., Foo, L. Y. & Ghaskadbi, S. S.** (2009). Radioprotective properties of polyphenols from *Phyllanthus amarus* Linn. *Journal of Radiation Research*, 50, 303-309.
- Lowry, J.B.** (1972). Anthocyanins of some Malaysian members of the *Gesneriaceae*. *Phytochemistry*, 11, 3267-74.
- Mansouri, M. T., Farbood, Y., Sameri, M. J., Sarkaki, A., Naghizadeh, B. & Rafeirad, M.** (2013). Neuroprotective effects of oral gallic acid against oxidative stress induced by 6-hydroxydopamine in rats. *Food Chemistry*, 138, 1028-1033.
- Markovska, Y., Kimenov, G., Stefanov, K. & Popov, S.** (1992). Lipid and sterol changes in leaves of *Haberlea rhodopensis* and *Ramonda serbica* at transition from biosis into anabiosis and vice versa caused by water stress. *Phytochemistry*, 31, 2309-2314.
- Materska, M., Konopacka, M., Rogoliński, J. & Ślosarek, K.** (2015). Antioxidant activity and protective effects against oxidative damage of human cells induced by X-radiation of phenolic glycosides isolated from pepper fruits *Capsicum annum* L. *Food Chemistry*, 168, 546-553.
- Maurya, D. K., Salvi, V. P. & Nair, C. K. K.** (2005). Radiation protection of DNA by ferulic acid under *in vitro* and *in vivo* conditions. *Molecular and Cellular Biochemistry*, 280, 209-217.
- Mihaylova, D. S., Bahchevanska, S. T. & Toneva, V. T.** (2011). Microwave assisted extraction of flavonoid antioxidant from leaves of *Haberlea Rhodopensis*. *Journal of international scientific publications: Materials, Methods & Technologies*, 5, 104-114.
- Moga, M., Dimienescu, O., Arvatescu, C., Mironescu, A., Dracea L. & Ples L.** (2016). The Role of Natural Polyphenols in the Prevention and Treatment of Cervical Cancer-An Overview. *Molecules*, 21, 1055-1087.
- Mori, H., Tanaka, T., Shima, H., Asu, T.K. & Takahashi, M.** (1986). Inhibitory effect of chlorogenic acid on methylazoxy-

- methanol acetate-induced carcinogenesis in large-intestine and liver of hamsters. *Cancer Letters*, 30, 49-54.
- Moyankova, D., Mladenov, P., Berkov, S., Peshev, D., Georgieva, D. & Djilianov, D.** (2014). Metabolic profiling of the resurrection plant *Haberlea Rhodopensis* during desiccation and recovery. *Physiologia Plantarum*, 152, 675-687.
- Mut-Salud, N., Álvarez, P. J., Garrido, J.M., Carrasco, E., Aránega, A., Rodríguez-Serrano, F.** (2016). Antioxidant Intake and Antitumor Therapy: Toward Nutritional Recommendations for Optimal Results. *Oxidative Medicine and Cellular Longevity*, ID 6719534, 1-19.
- Nagao, A., Seki, M. & Kobayashi, H.** (1999). Inhibition of xanthine oxidase by flavonoids. *Bioscience, Biotechnology, and Biochemistry*, 63, 1787-1790.
- Nair, G.G. & Nair, C. K. K.** (2013). Radioprotective Effects of Gallic Acid in Mice. *BioMed Research International*, 2013, 1-13.
- Özyurt, H., Çevik, Ö., Özgen, Z., Özden, A.S., Çadırıcı, S., Elmas, M.A., Ercan, F., Gören, M.Z. & Şener, G.** (2014). Quercetin protects radiation-induced DNA damage and apoptosis in kidney and bladder tissues of rats. *Free Radical Research*, 48, 1247-1255.
- Patil, S. L., Mallaiah, S.H. & Patil, R. K.** (2013). Antioxidative and radioprotective potential of rutin and quercetin in Swiss albino mice exposed to gamma radiation. *Journal of Medical Physics*, 38, 87-92.
- Patil, S. L., Rao, N. B., Somashekarappa, N. M. & Rajashekhar, K.P.** (2014). Antigenotoxic Potential of Rutin and Quercetin in Swiss Mice Exposed to Gamma Radiation. *Biomedical Journal*, 37, 305-313.
- Popov, B., Georgieva, S. & Lalchev, S.** (2012). Radioprotection from genetic damages by resurrection plant *Haberlea rhodopensis* – in vivo/in vitro study with rabbits. *Trakia Journal of Science*, 10, 41-49.
- Popov, B., Georgieva, S., Gadjeva, V. & Petrov, V.** (2011). Radioprotective, anticlastogenic and antioxidant effects of total extract of *Haberlea Rhodopensis* on rabbit blood samples exposed to gamma radiation in vitro. *Revue de Medecine Veterinaire*, 162, 34-39.
- Popov, B., Georgieva, S., Oblakova, M. & Bonev, G.** (2013). Effects of *Haberlea rhodopensis* extract on antioxidation and lipid peroxidation in rabbits after exposure to <sup>60</sup>Co-γ-rays. *Arch Biol Sci Belgrade*, 65, 91-97.
- Popov, B., Radev, R. & Georgieva, S.** (2010). In vitro incidence of chromosome aberrations in gamma-irradiated rabbit lymphocytes, treated with *Haberlea rhodopensis* extract and vitamin C". *Bulgarian Journal of Veterinary Medicine*, 13, 148-153.
- Pradeep, K., Park, S. H. & Ko, K. C.** (2008). Hesperidin a flavanoglycone protects against γ- irradiation induced hepatocellular damage and oxidative stress in Sprague–Dawley rats. *European Journal of Pharmacology*, 587, 273-280.
- Prasad, N.R., Srinivasan, M., Pugalendi, K.V. & Menon, V.P.** (2006). Protective effect of Ferulic acid on γ-radiation induced micronuclei, dicentric aberration and lipid peroxidation in human lymphocytes. *Environmental Mutagenesis*, 603, 129-134.
- Rehmana, M. U., Jawaida, P., Uchiyamab, H. & Kondo, T.** (2016). Comparison of free radicals formation induced by cold atmospheric plasma, ultrasound, and ionizing radiation. *Archives of Biochemistry and Biophysics*, 605, 19-25.
- Ross, J.A. & Kasum, C. M.** (2002). Dietary flavonoids: bioavailability, metabolic effects, and safety. *Annual Review of Nutrition*, 22, 19-34.
- Sancar, A., Lindsey-Boltz, L. A., Unsal-Kaçmaz, K. & Linn, S.** (2004). Molecular mechanisms of mammalian DNA repair and the DNA damage checkpoints. *Annual Review of Biochemistry*, 73, 39-85.
- Semwal, D.K., Semwal, R. B., Combrinck, S. & Viljoen, A.** (2016). Myricetin: A Dietary Molecule with Diverse Biological Activities. *Nutrients*, 8, 90-121.
- Sevgi, K., Tepe, B. & Sarikurkcu, C.** (2015). Antioxidant and DNA damage protection potentials of selected phenolic acids. *Food and Chemical Toxicology*, 77, 12-21.
- Shaikh, S. & Shah, M.** (2018). Radio-protective potential and antimicrobial activity of pudina. (*Mentha spp.* /mint). *World Journal of Pharmaceutical Research*, 7, 1735-1752.
- Shanthakumar, J., Karthikeyan, A., Bandugula, V.R. & Prasad, N.R.** (2012). Ferulic acid, a dietary phenolic acid, modulates radiation effects in Swiss albino mice. *European Journal of Pharmacology*, 691, 268-274.
- Shen, Y., Song, X., Li, L., Sun, J., Jaiswal, Y., Huang, J., Liu, C., Yang, W., Williams, L., Zhang, H. & Guan, Y.** (2019). Protective effects of p-coumaric acid against oxidant and hyperlipidemia-an in vitro and in vivo evaluation. *Biomedicine and Pharmacotherapy*, 111, 579-587.
- Shimoi, K., Masuda, S., Shen, B., Furugori, M. & Kinae, N.** (1996). Radioprotective effects of antioxidative plant flavonoids in mice. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 350, (1) 153-161.
- Silva, J. P., Gomes, A. C. & Coutinho, O. P.** (2008). Oxidative DNA damage protection and repair by polyphenolic compounds in PC12 cells. *European Journal of Pharmacology*, 601, 50-60.
- Simões, G. D., Hecktheuer, L. H. R., Hubscher, G.H. & Boligon, A. A.** (2018). Quantification of bioactive compounds in crem (*Tropaeolum pentaphyllum* Lam) tubers: fibers, phenolic compounds and evaluation of its antioxidant activity. *International Food Research Journal*, 25(3), 1315-1321
- Sova, M.** (2012). Antioxidant and antimicrobial activities of cinnamic acid derivatives. *Mini reviews in medicinal chemistry*, 12, 749-767.
- Sricharan, P., Lamaiphan, N., Patthawaro, P., Limchoowong, N., Techowongstien, S. & Chanthai, S.** (2017). Phytochemicals in Capsicum oleoresin from different varieties of hot chilli peppers with their antidiabetic and antioxidant activities due to some phenolic compounds. *Ultrasonics Sonochemistry*, 38, 629-639.
- Srinivasan, M., Sudheer, A.R. & Menon, V. P.** (2007). Ferulic Acid: Therapeutic potential through its antioxidant property. *Journal of Clinical Biochemistry and Nutrition*, 40, 92-100.
- Sun, F., Zheng, X. Y., Ye, J., Wu, T. T., Wang, J. & Chen, W.** (2012). Potential anticancer activity of myricetin in human T24 bladder cancer cells both in vitro and in vivo. *Nutrition and Cancer*, 64, 599-606.
- Taheri, A., Rostamzadeh, A., Gharib, A., & Fatehi, D.** (2016). Radioprotective effects of Silymarin, a natural medical herb, in modulation and prevention of radiation induced damages.

- Scholars Research Library, Der Pharmacia Lettre*, 8, 146-150.
- Thiyam, U., Stöckmann, H., Felde, T.Z. & Schwarz, K.** (2006). Antioxidative effect of the main sinapic acid derivatives from rapeseed and mustard oil by-products. *European Journal of Lipid Science and Technology*, 108, 239-48.
- Van Buren, R., Wai, C. M., Giarola, V., Ambrosini, S., Song, X. & Bartels, D.** (2018). Desiccation tolerance evolved through gene duplication and network rewiring in *Lindernia*. *The Plant Cell*, 30, 2943-2958.
- Verma, M., Rai, G.K. & Kaur, D.** (2018). Effect of extraction solvents on phenolic content and antioxidant activities of Indian gooseberry and guava. *International Food Research Journal*, 25(2), 762-768.
- Verma, S., Singh, A. & Mishra, A.** (2013). Gallic acid: Molecular rival of cancer. *Environmental Toxicology and Pharmacology*, 35, 73-485.
- Zbikowska, H. M., Szejka, M., Saluk, J., Pawlaczyk-Graja, I., Gancarz, R. & Olejnik, A. K.** (2016). Polyphenolic-polysaccharide conjugates from plants of Rosaceae/Asteraceae family as potential radioprotectors. *International Journal of Biological Macromolecules*, 86, 329-337.
- Zheleva-Dimitrova, D., Nedialkov, P. & Giresser, U.A.** (2016). Validated HPLC Method for Simultaneous Determination of Caffeoyl Phenylethanoid Glucosides and Flavone 8-C-glycosides in *Haberlea rhodopensis*. *Natural Product Communications*, 11, 791-792.